

Biotechnology for Recovery of Rare Metals in Wastewater



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Rare-metal Bioremediation and Recycling using Biotechnology



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AGENDA

Selenium as a resource and a pollutant

Double benefit concept for selenium recovery from wastewater

Selenium-reducing bacterium (1)

Bacillus selenatarsenatis SF-1

Selenium-reducing bacterium (2)

Pseudomonas stutzeri NT-1

Possible applications of metal biology to other rare metals

Rare Metals

Group	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
Period																		
1	1 H																	2 He
2	3 Li	4 Be											5 B	6 C	7 N	8 O	9 F	10 Ne
3	11 Na	12 Mg											13 Al	14 Si	15 P	16 S	17 Cl	18 Ar
4	19 K	20 Ca	21 Sc	22 Ti	23 V	24 Cr	25 Mn	26 Fe	27 Co	28 Ni	29 Cu	30 Zn	31 Ga	32 Ge	33 As	34 Se	35 Br	36 Kr
5	37 Rb	38 Sr	39 Y	40 Zr	41 Nb	42 Mo	43 Tc	44 Ru	45 Rh	46 Pd	47 Ag	48 Cd	49 In	50 Sn	51 Sb	52 Te	53 I	54 Xe
6	55 Cs	56 Ba	* 71 Lu	72 Hf	73 Ta	74 W	75 Re	76 Os	77 Ir	78 Pt	79 Au	80 Hg	81 Tl	82 Pb	83 Bi	84 Po	85 At	86 Rn
7	87 Fr	88 Ra	** 103 Lr	104 Rf	105 Db	106 Sg	107 Bh	108 Hs	109 Mt	110 Ds	111 Rg	112 Uub	113 Uut	114 Uuq	115 Uup	116 Uuh	117 Uus	118 Uuo

*Lanthanoids	* 57 La	58 Ce	59 Pr	60 Nd	61 Pm	62 Sm	63 Eu	64 Gd	65 Tb	66 Dy	67 Ho	68 Er	69 Tm	70 Yb
**Actinoids	** 89 Ac	90 Th	91 Pa	92 U	93 Np	94 Pu	95 Am	96 Cm	97 Bk	98 Cf	99 Es	100 Fm	101 Md	102 No

 Rare Metal
 Rare Earth

Consumption of metals is increasing in hi-tech Industries.

Environmental Issues Related to Rare Metals

Rare metals are emitted into environment via wastes and wastewater



Environmental Contamination

Elevated concentration of rare metals can cause toxic or adverse effects on the environment.

Decay/Depletion as Resource

Shortage as resource has started due to loss as wastewater and wastes, soaring market price of elements.



Recovery of rare metals from wastes/wastewater is necessary for sustainable society.

Selenium as Example

As Environmental Contaminant

Se-Wastewater

Selenate (SeO_4^{2-}) : Soluble Se

Selenite (SeO_3^{2-}) : **Toxic**

Environmental standard on water quality $\leq 0.01\text{mg-Se/L}$

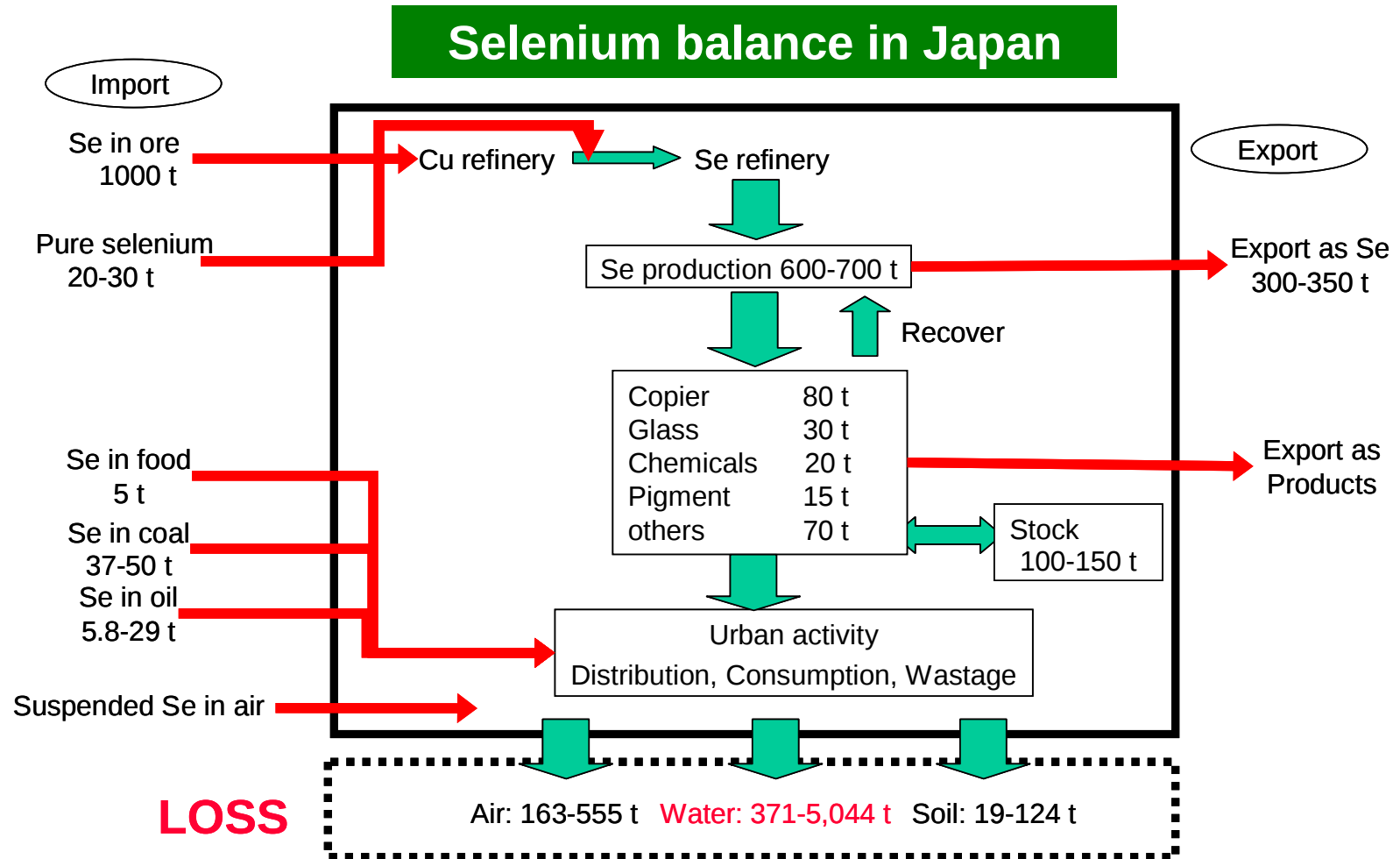
Regulation of industrial wastewater $\leq 0.1\text{mg-Se/L}$

Provisional regulation for Se industry $\leq 0.3 \text{ mg-Se/L}$

Industry	Se conc.
Oil refinery	50-300 $\mu\text{g/l}$
Copper purification plant	>20 mg/l
Chemical plant	250 mg/l

Selenium as Example

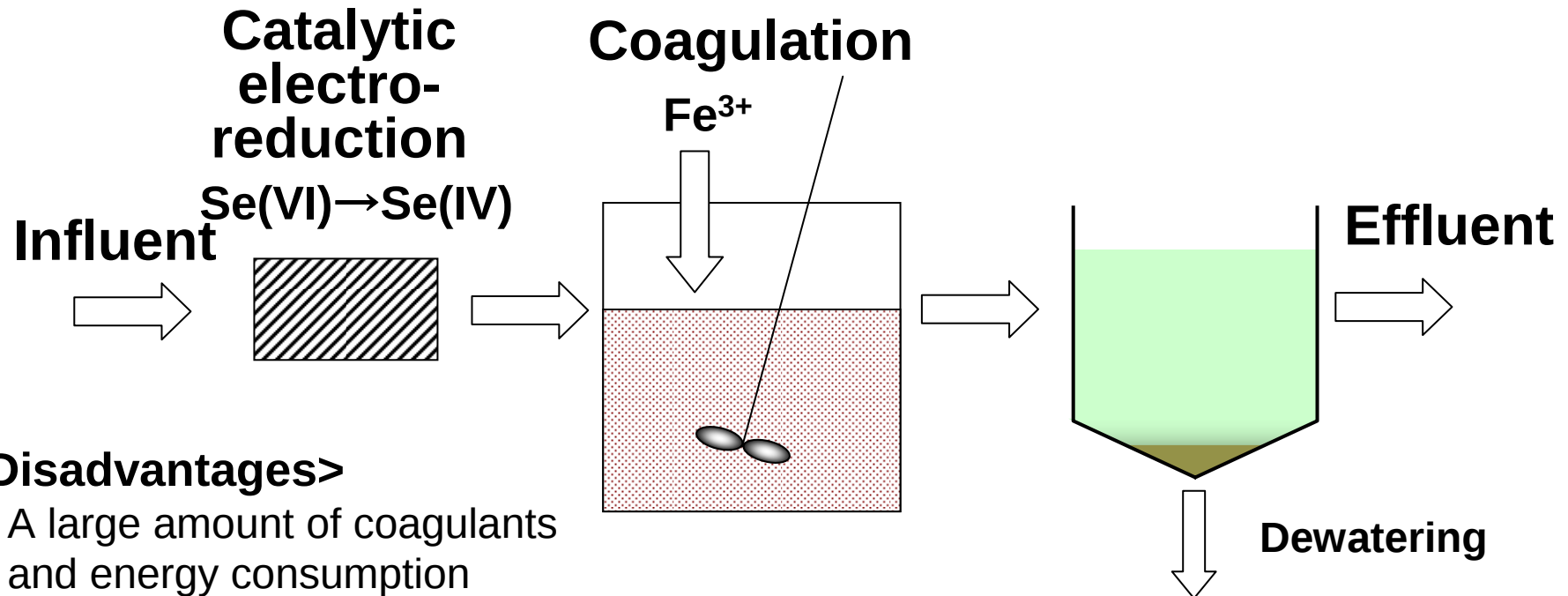
As Resource



Market Price: 2.01 \$/lb in 1996 ➡ 47.4 \$/lb in 2005

Existing Se Wastewater Treatment

Physico-Chemical Process



<Disadvantages>

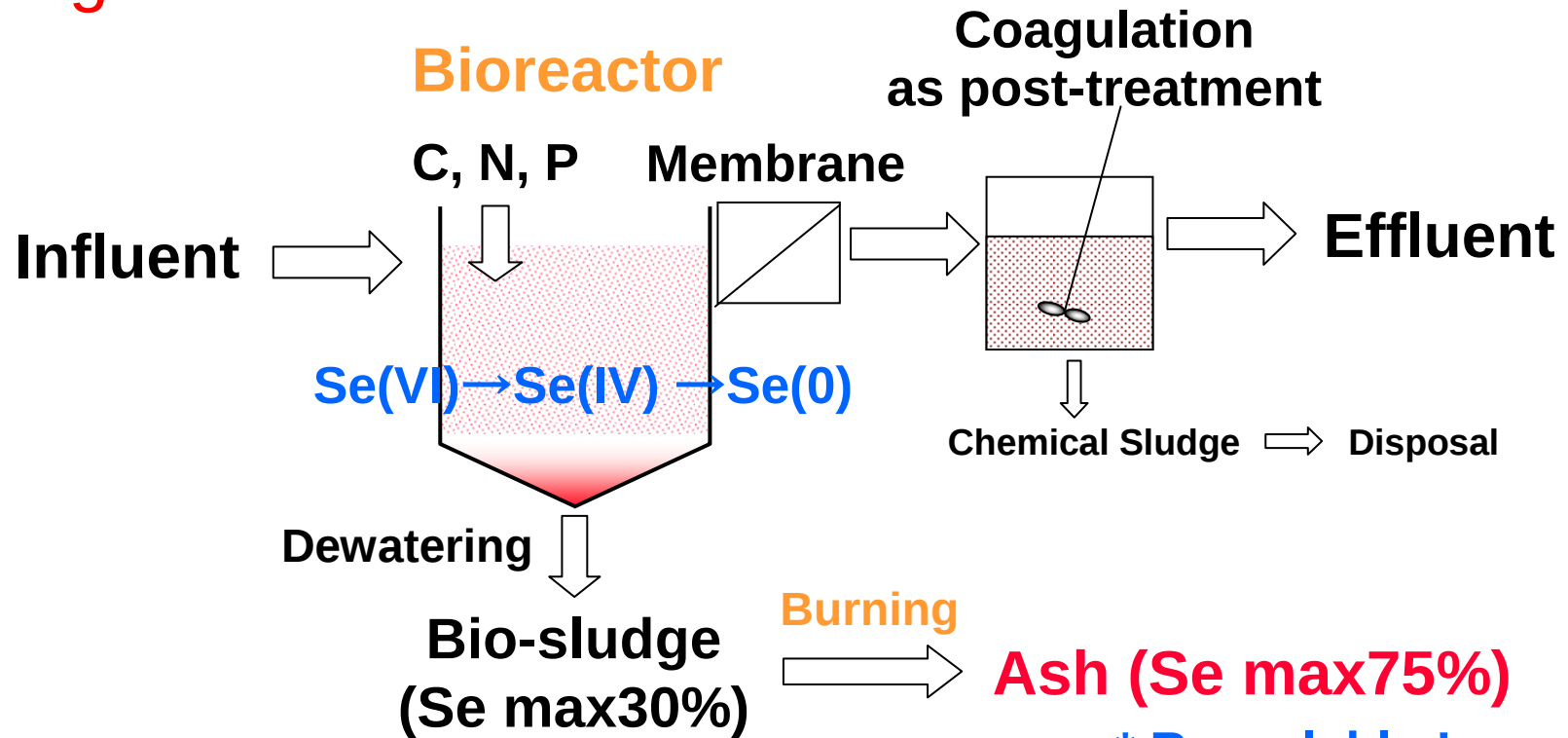
- A large amount of coagulants and energy consumption
- Generation of a large amount of sludge
- Low resource value of sludge due to low Se content
- High-cost (including waste disposal cost)

Chemical sludge (Se <1%)
* Cannot be recycled due to low Se content

**Waste disposal cost
¥30,000/ton-sludge!**

Novel Concept for Se Recovery from Wastewater

Biological Process



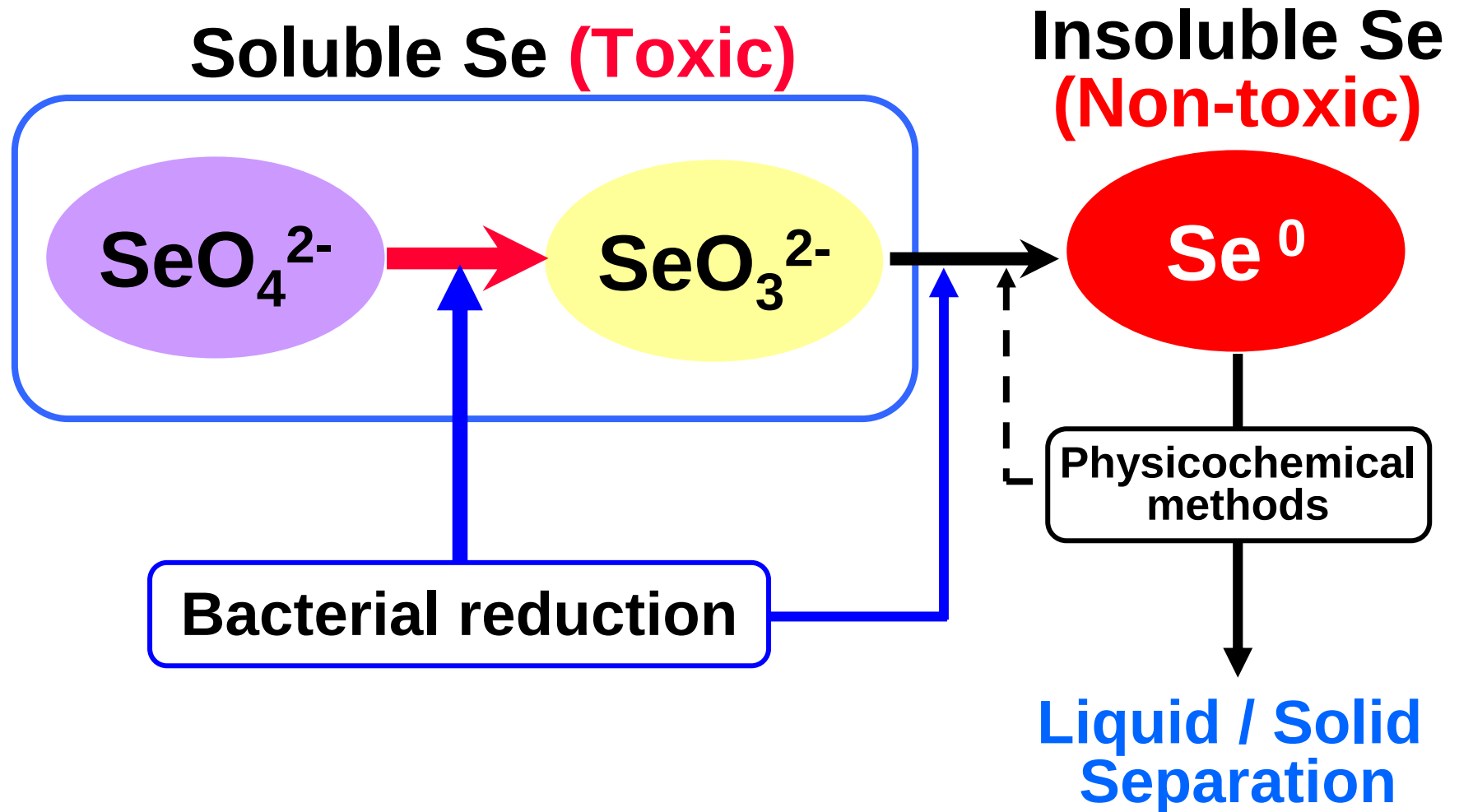
<Advantages>

- Low energy and reagent consumption
- Low sludge generation
- High resource value of bio-sludge due to high Se content
- Disposal cost turns a profit by Se recycle

*** Recyclable !**
Due to high Se content
Benefit ¥ 650,000/ton-ash

Key Microbial Reaction

Selenate/Selenite Reduction into Elemental Se



Selenate-Reducing Bacterium (1)

Bacillus selenatarsenatis SF-1

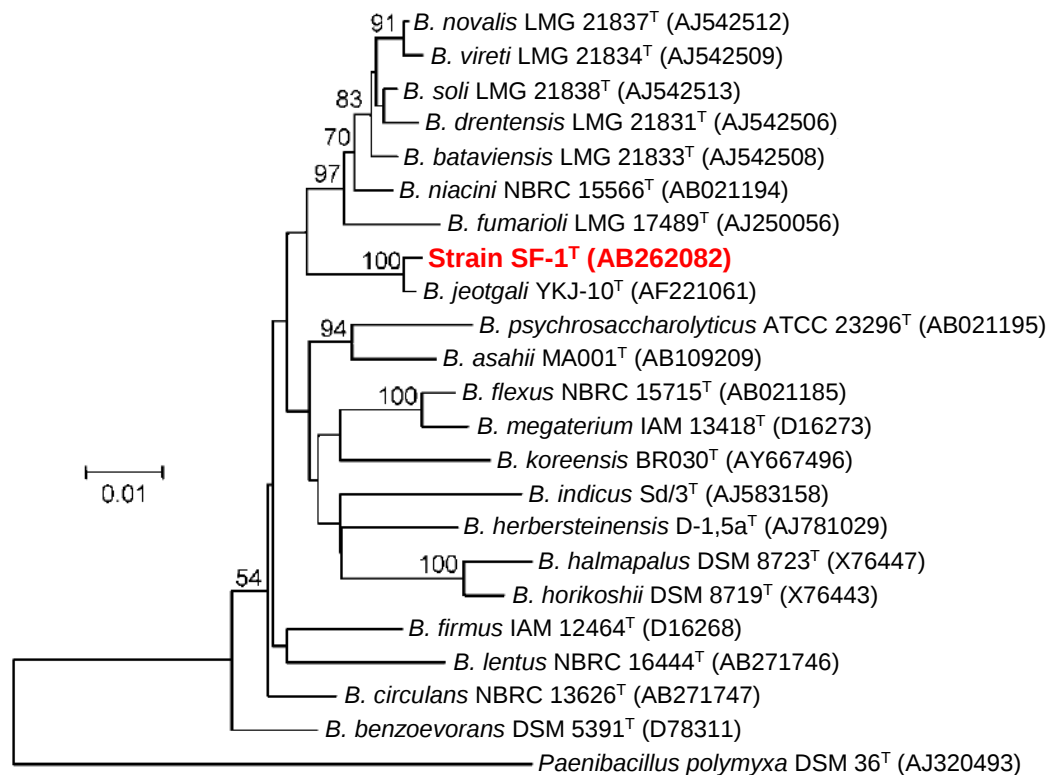
Origin: Se-contaminated sediment

Taxonomy: gram positive, facultative anaerobe

Selenate reduction: respiration with selenate as e^- acceptor



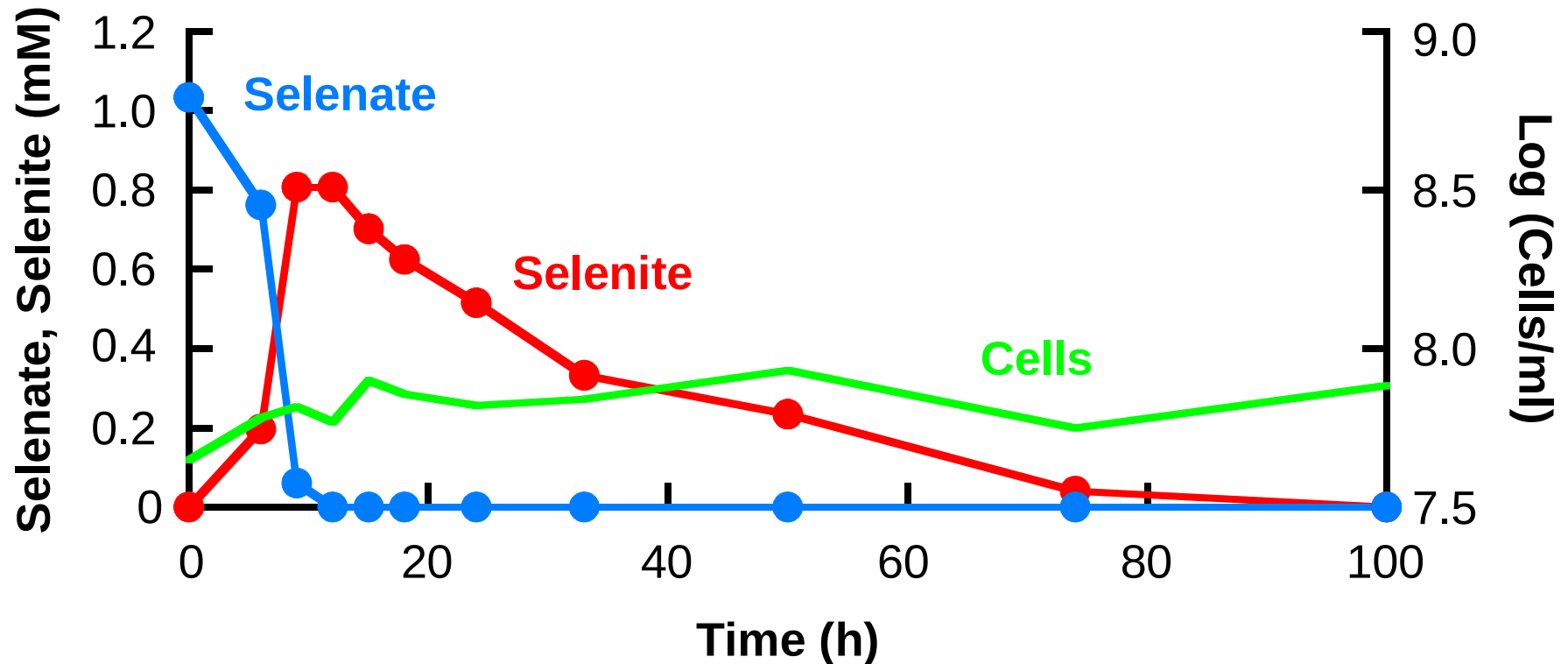
Novel species of *Bacillus*
Bacillus selenatarsenatis



Fujita et al. (1997) J. Ferment. Bioeng., 83, 517-522;.

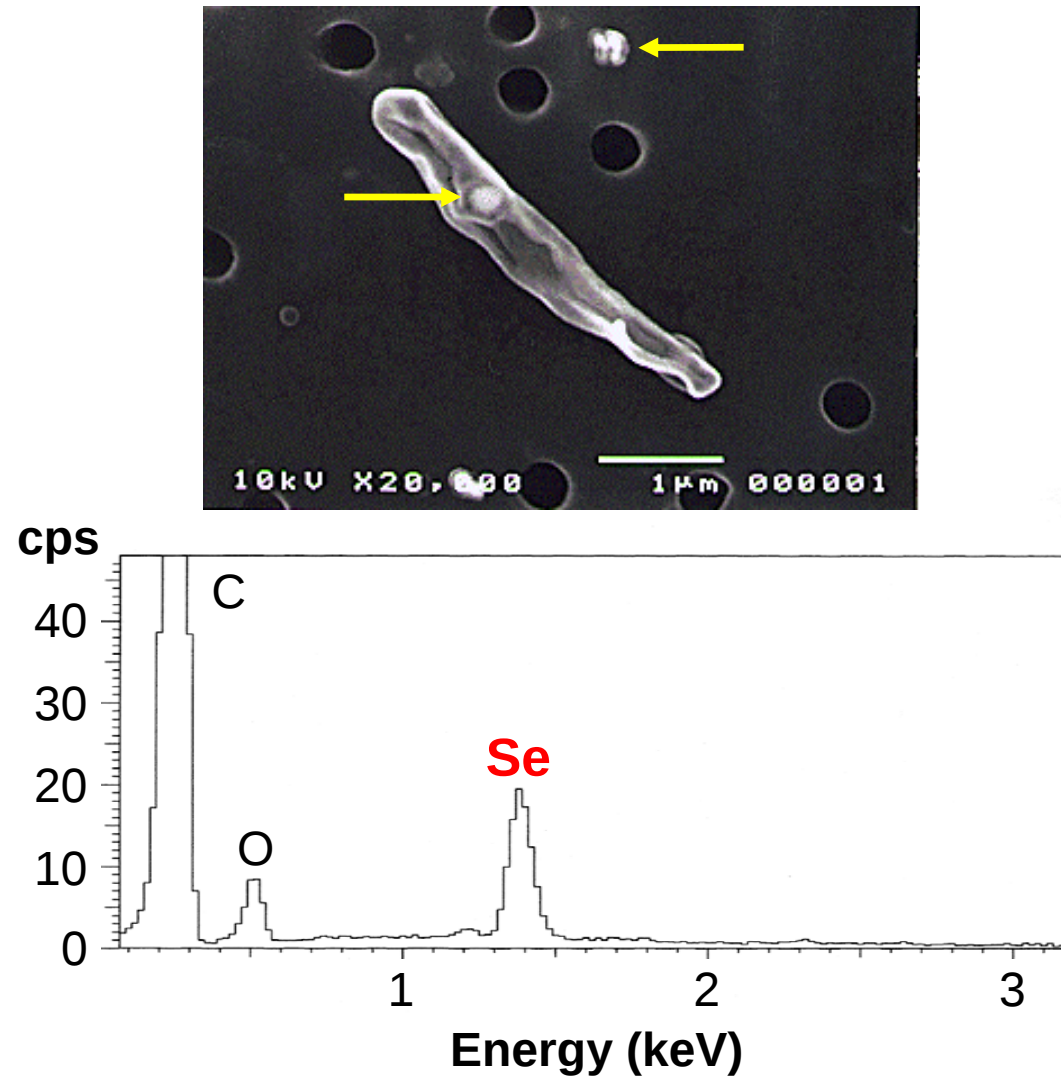
Yamamura et al. (2007) Int. J. Syst. Evol. Microbiol., 57, 1060-1064.

Selenate Reduction by *B. selenatarsenatis* SF-1



Under anoxic condition (energy saving)
Rate-determining step in selenite reduction

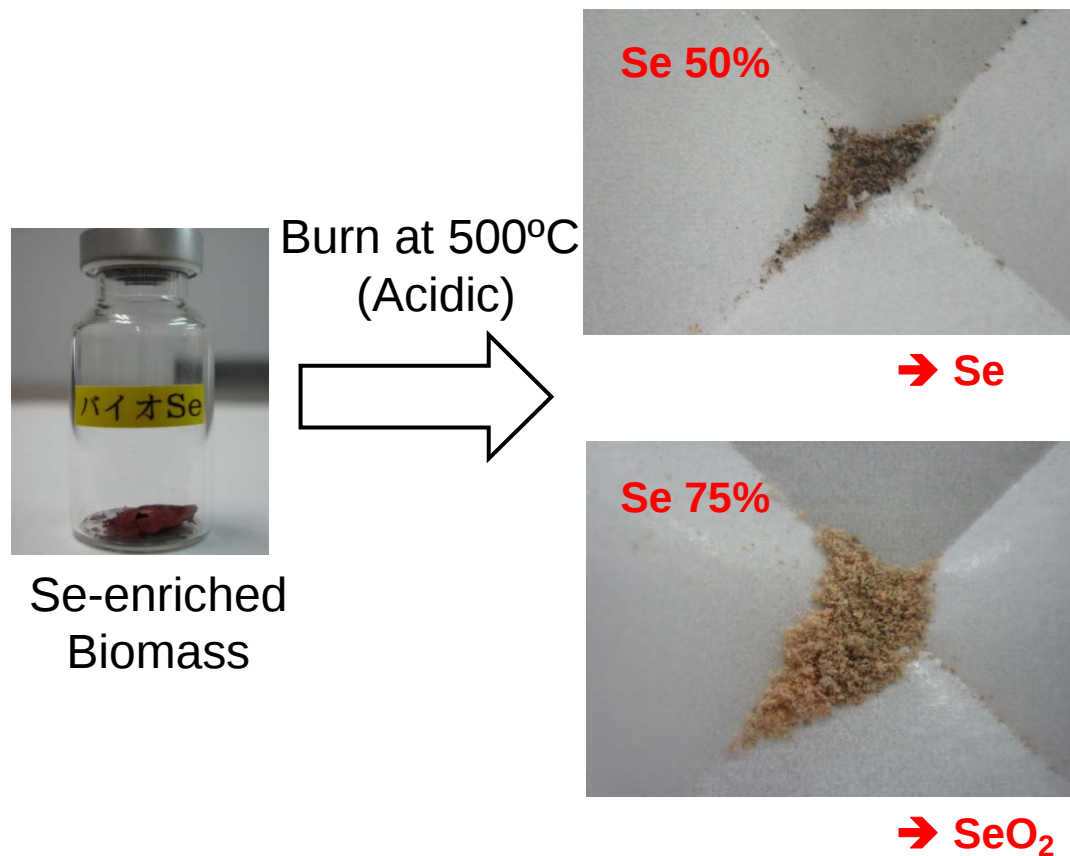
Deposition of Elemental Se by *B. selenatarsenatis*SF-1



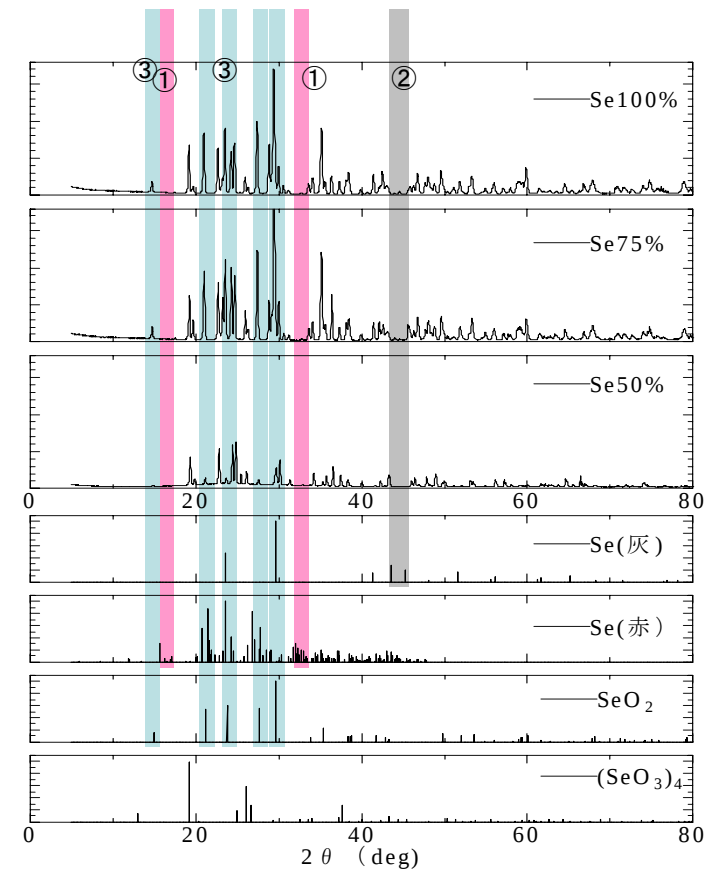
Se deposition in cells (need for extraction from biomass)

Lab-scale Se Recovery Process with *B. selenatarsenatis*SF-1

Se Recovery from Biomass



X-Ray Diffraction (XRD) analysis



Se metal or SeO₂ can be recovered from the biomass

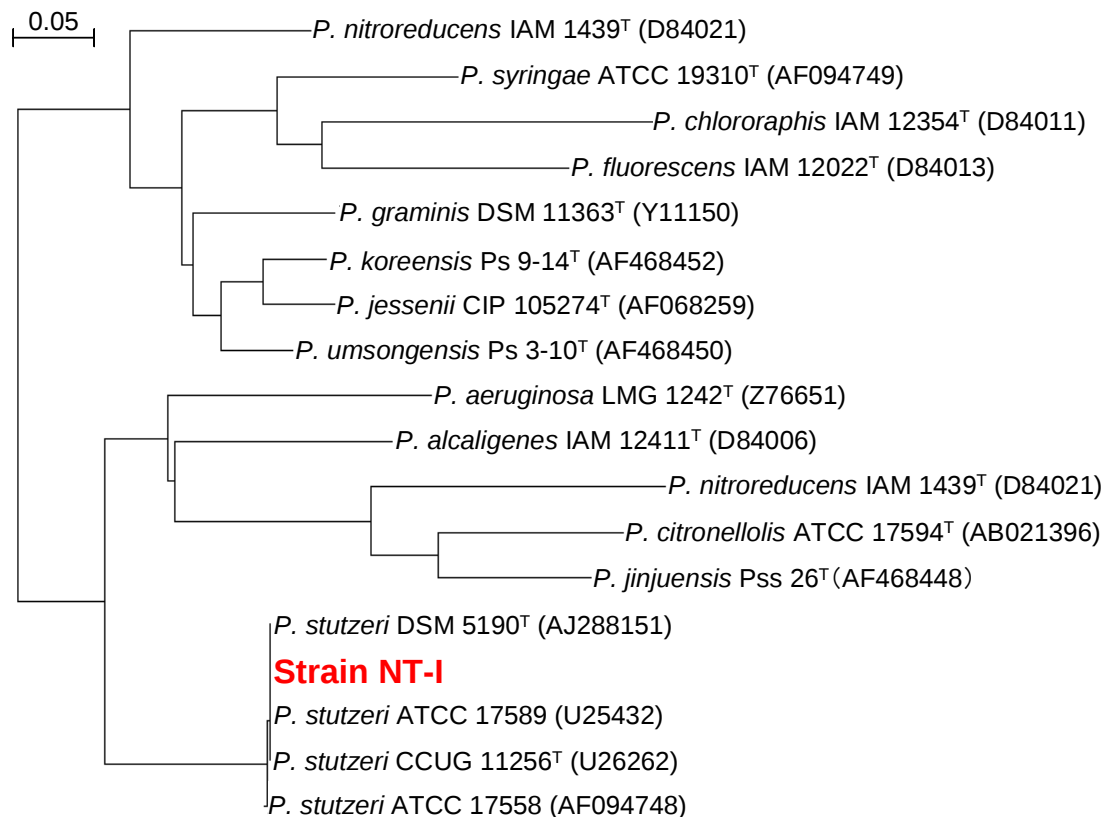
Selenate-Reducing Bacterium (2)

Pseudomonas stutzeri NT- I

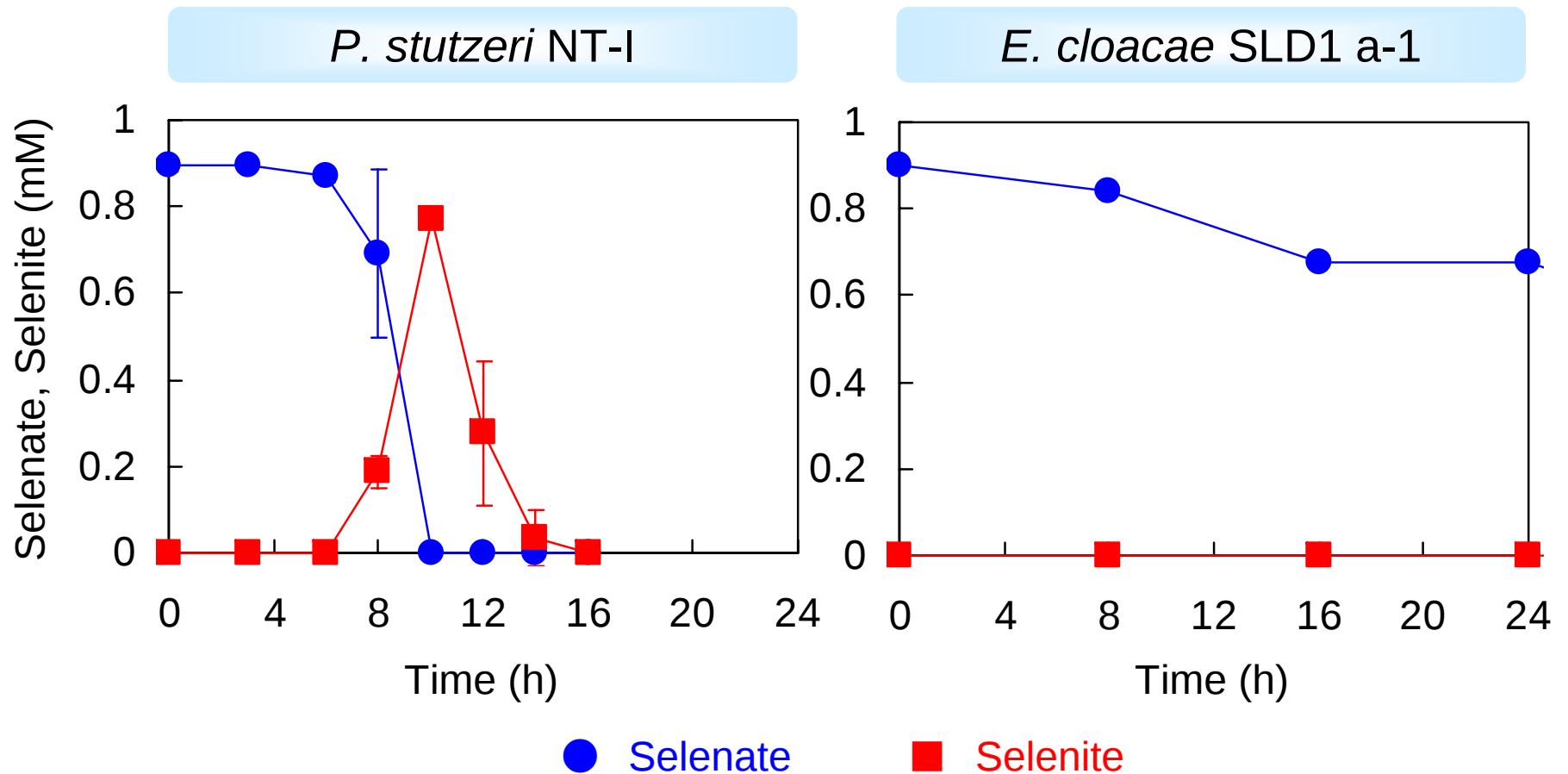
Origin: Biofilm from Se wastewater drainage

Taxonomy: gram negative, aerobe

Selenate reduction: in the presence of oxygen (detoxification?)

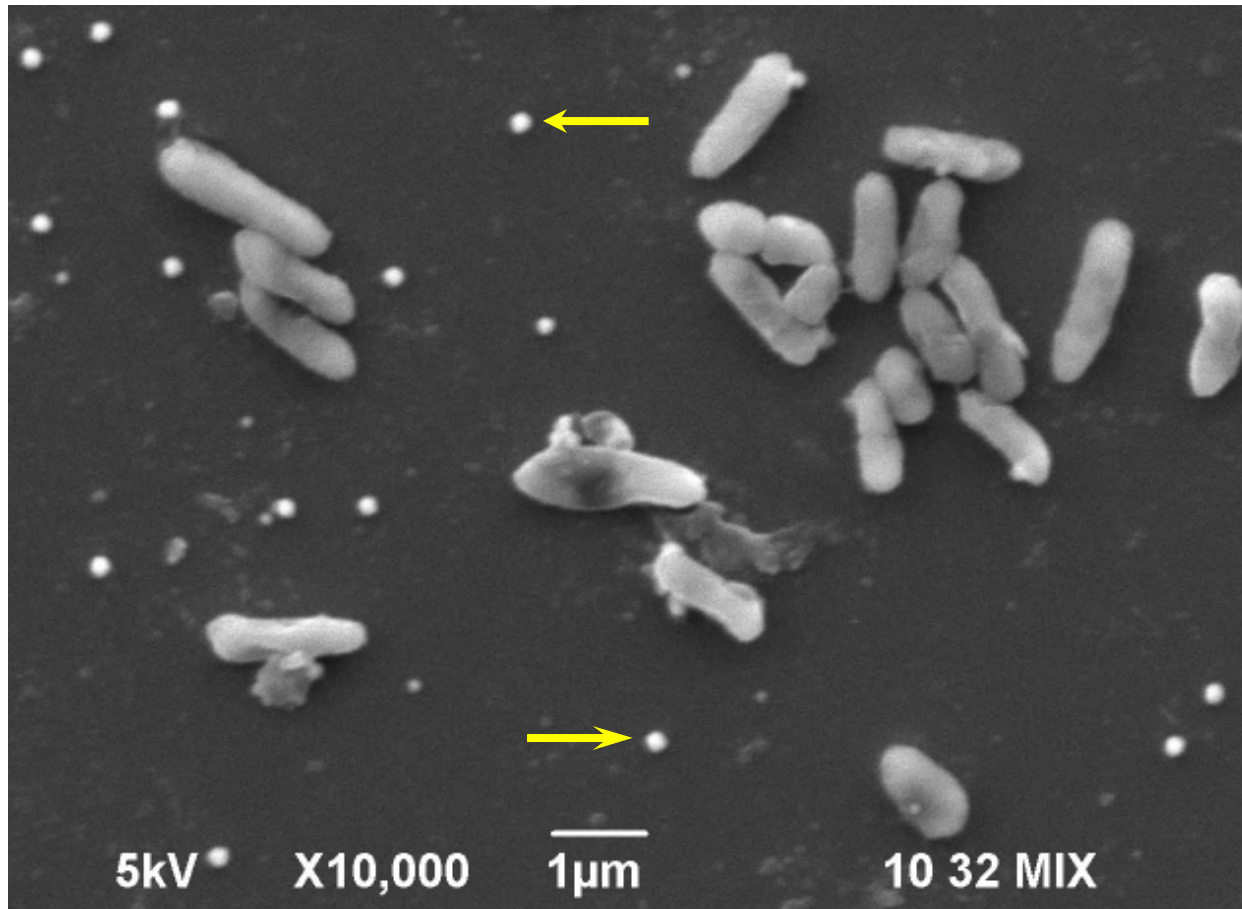


Selenate Reduction Property by *P. stutzeri* NT- I



Under aerobic condition (energy consuming)
Very effective reduction (the fastest reduction rate reported)

Deposition of Elemental Se by *P. stutzeri* NT- I



Se deposition outside cells (simple separation may be possible)

Pilot-scale Se Recovery Process with *P. stutzeri* NT- I

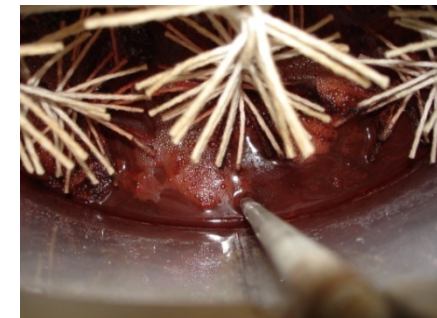
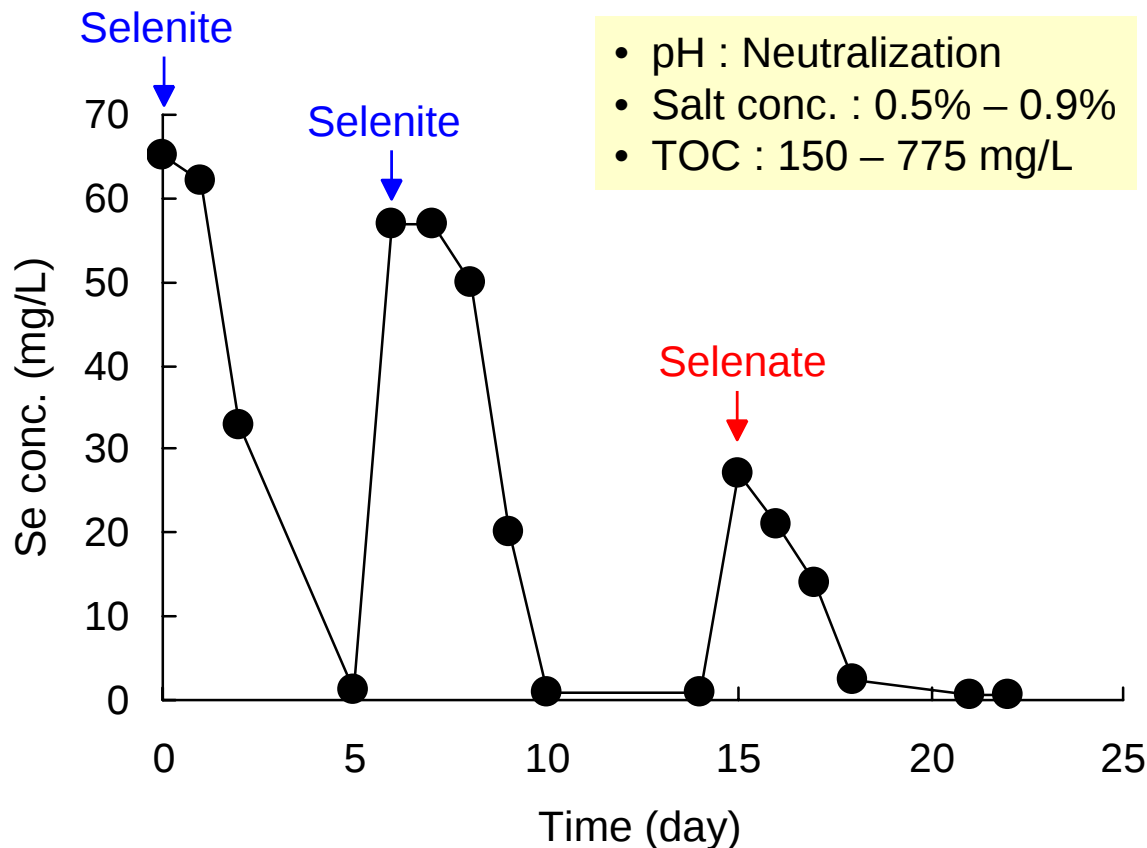
Pilot-scale Reactor at Shinko Chemical Co Ltd.



Reactor Overview (0.2 million yen for 2 reactors, 200L × 2) UASB

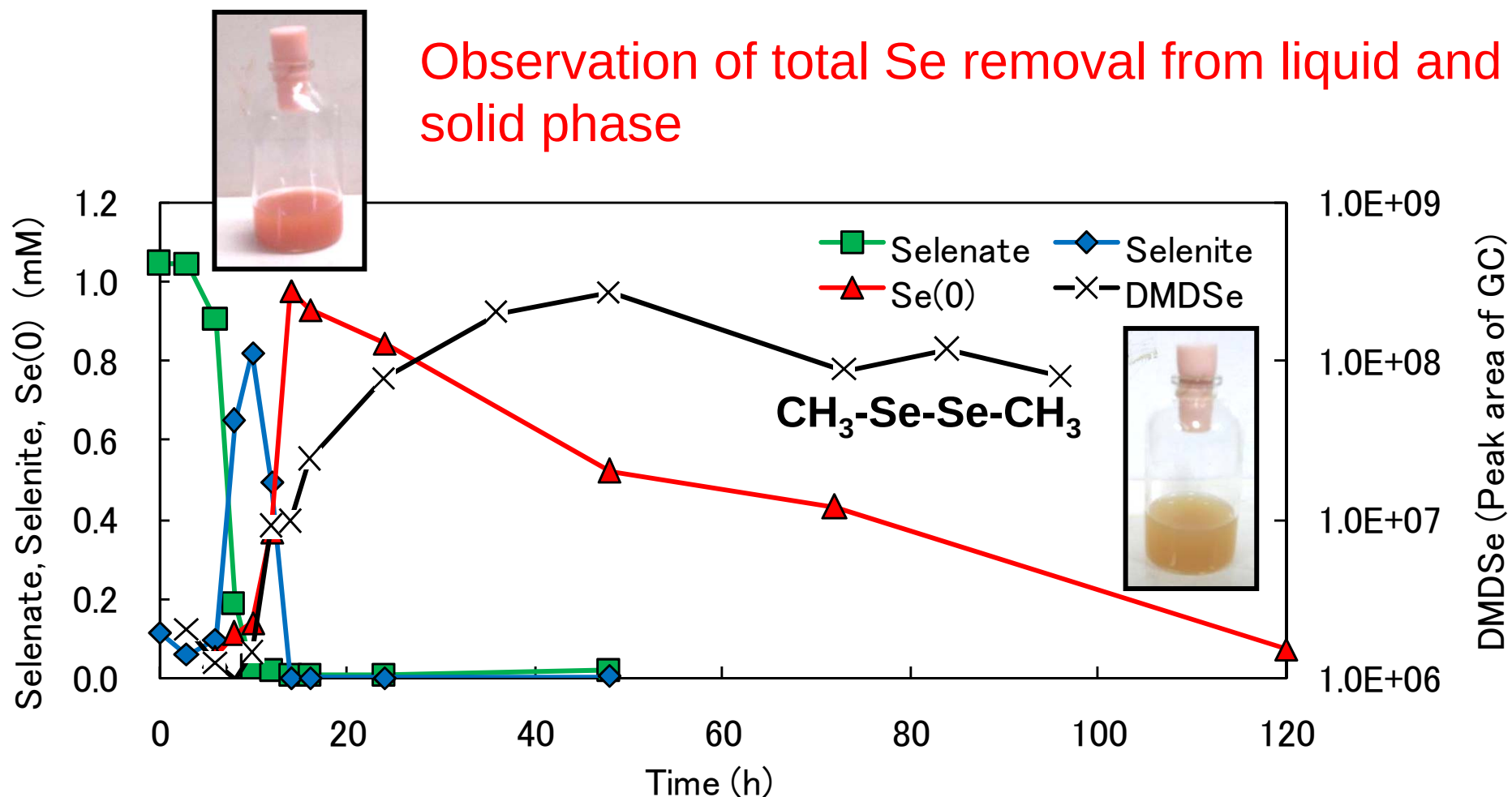
Pilot-scale Se Recovery Process with *P. stutzeri* NT-

Sequential batch trial using real industrial wastewater with NT-I attached onto biomass carrier



Efficient removal of soluble Se is achieved with weak aeration.

Possibility of Se Recovery by Volatilization (DMDSe)



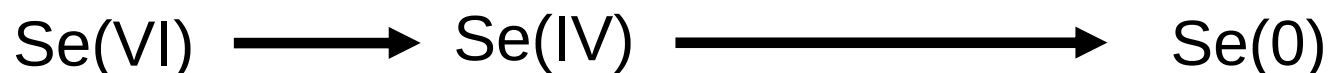
More cost-effective and simple Se recovery ?

Possible Application to Other Rare Metals

Transformation to Insoluble or Less-soluble Forms by
Reduction of Metal Oxyanions

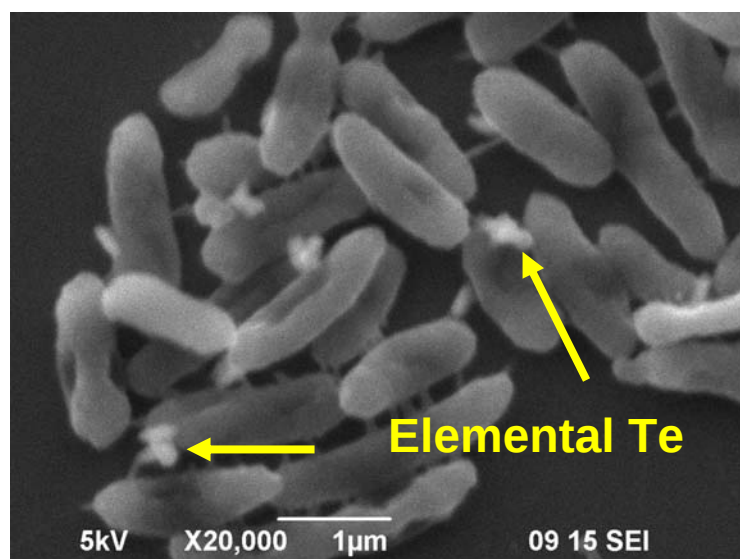
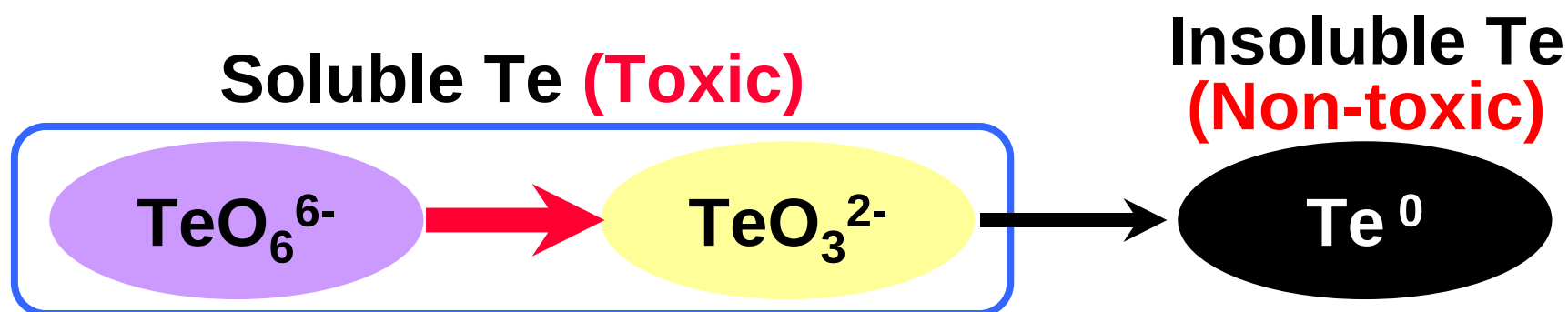
Soluble

Insoluble/less-soluble

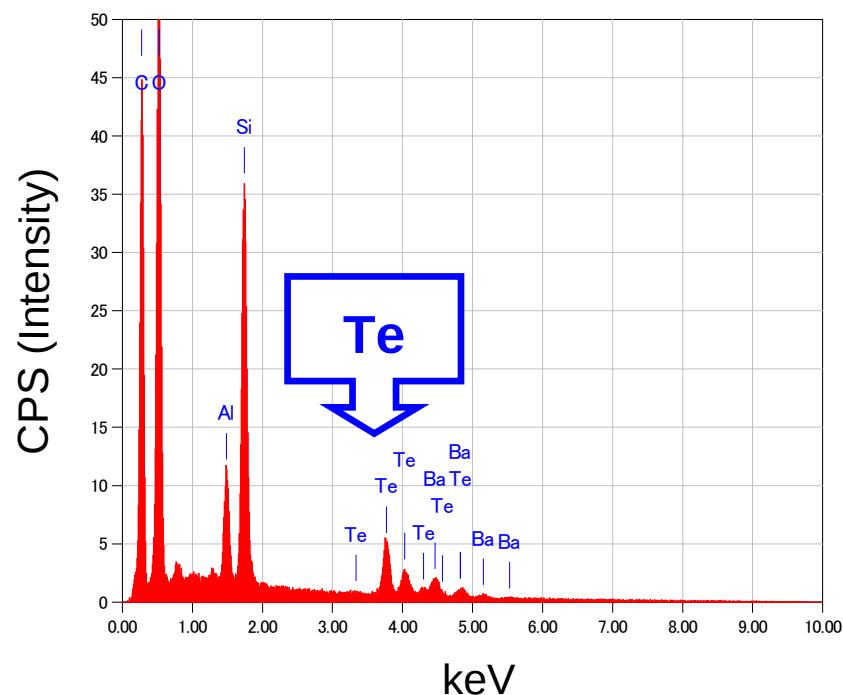


Possible Application to Other Rare Metals

Screening of Tellurate/Tellurite-Reducing Bacteria



Stenotrophomonas maltophilia TI-1



Possible Applications of Metal-Biotechnology



Wastewater treatment
for metal-containing
wastewater



Bioremediation
for polluted wetlands



Bioremediation
for polluted soils

Double Benefit Metal-Biotechnology

Metal recovery, recycling, processing

Low energy consumption, Speed up, New pathways, New compound (nano-particles), etc.

“Metal-Biotechnology” for Conservation of Environment and Resource

Bioprocess including reduction of metal-oxyanions has a great potential for concentration, extraction, and recover of rare metals from wastewater, wastes, and natural environment in cost-effective and eco-harmonizing manner.

Metal bioprocess not only in wastewater treatment and resource recovery (end-of-pipe) but also in mining, production, and processing (upper-stream application) can establish sustainable metal industries for future.



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FOUNDATION

Regional New Consortium Project, the Ministry of Economy, Trade and Industry, Japan, the Kansai Bureau Economy, Trade and Industry (2007)

Feasibility Study, Innovation Plaza Osaka, Japan Science and Technology Agency (2006)

Biotechnology for Recovery of Rare Metals in Wastewater

■ INVESTMENT FOR A TRIAL PRODUCT FOR A SPECIFIC MARKET

1. Selenium: treatment & recovery:

Investment fee : About \$ 0.4 Million

(Royalty, any cost for patent is not included)

Period : 3 years

2. Other metals: treatment & recovery

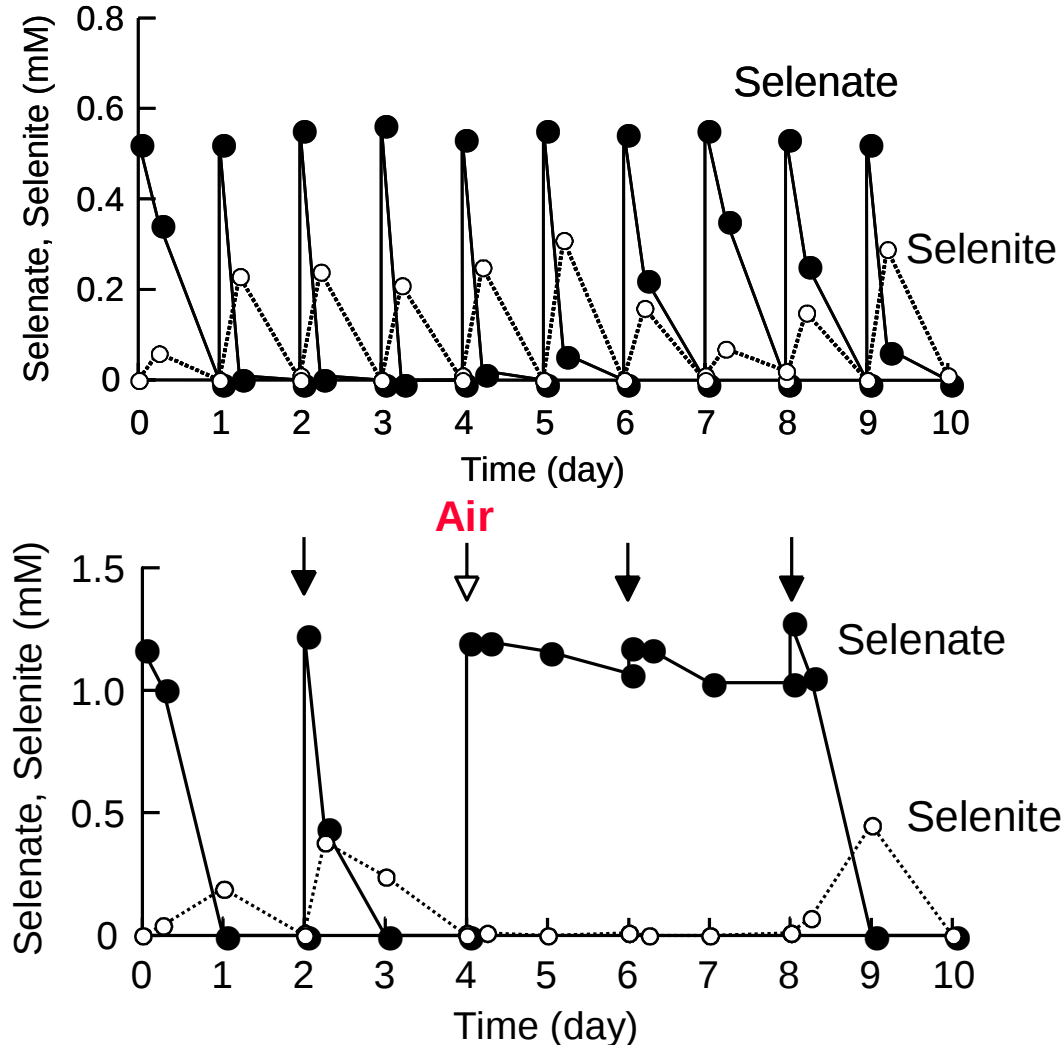
Investment fee : About \$ 0.8 Million

(Royalty, any cost for patent is not included)

Period : 5 years

Lab-scale Se Recovery Process with *B. selenatarsenatis*SF-1

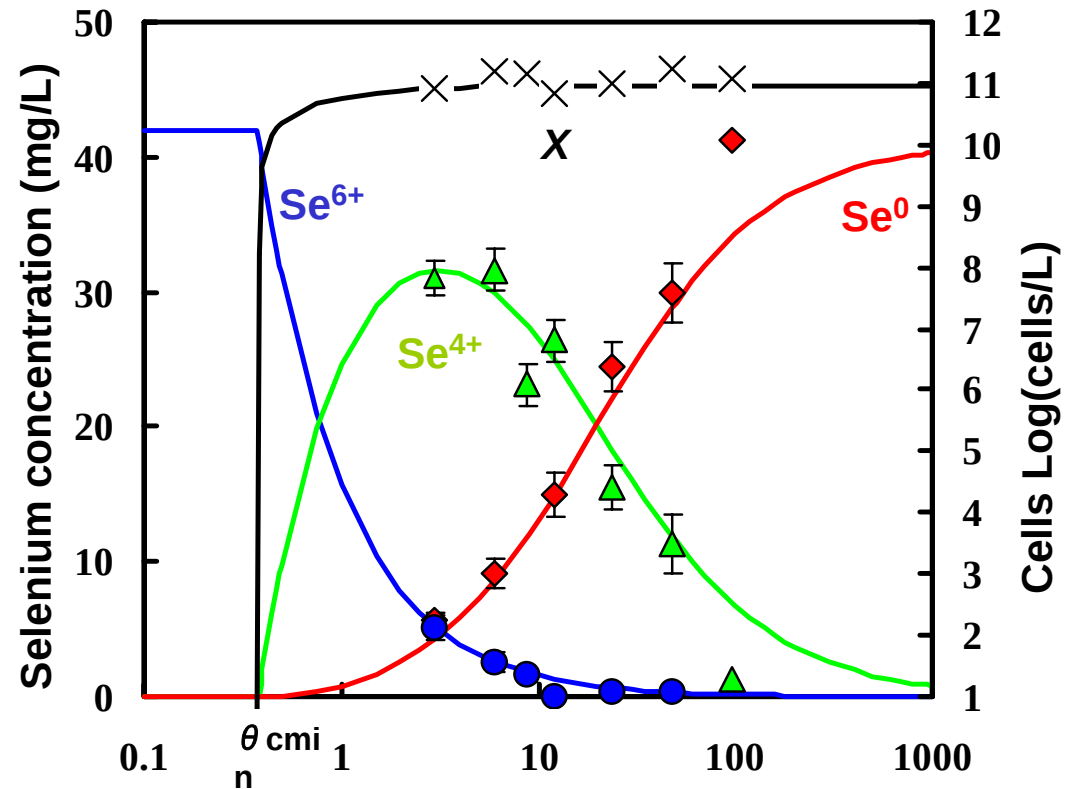
Sequential Batch Tests



Strict control of anoxic condition is required.

Lab-scale Se Recovery Process with *B. selenatarsenatis* SF-1

Continuous Flow Tests



Kinetics of the reactor

$$[Se^{6+}] = \frac{1}{k_1 Y_1 \theta_c}$$

$$[Se^0] = \frac{k_2 (k_1 Y_1 [Se^{6+}]_{in} \theta_c - 1)^2}{k_1 Y_1 \theta_c (k_1 - k_2 + k_1 k_2 Y_1 [Se^{6+}]_{in} \theta_c)}$$

Cell retention time θ_c
(h)

$$[Se^{4+}] = \frac{k_1 Y_1 [Se^{6+}]_{in} \theta_c - 1}{Y_1 \theta_c (k_1 - k_2 + k_1 k_2 Y_1 [Se^{6+}]_{in} \theta_c)}$$

$$[X] = Y_1 [Se^{6+}]_{in} - \frac{1}{k_1 \theta_c}$$

Rare-metal Bioremediation and Recycling using Biotechnology

Mitsuo YAMASHITA

Shibaura Institute of Technology

Faculty of Engineering

Rare metals

○ a few resource, ○ hard to mine and refine in technical and economical, ○ a rapid increasing in use depending on the development of high technology such as IT, Space, Car, Energy Industry, ○ harmful contaminant

Possibility of Depleting Resource

Expensive in market, nation's reservation:

Ga, Ge, Se, Te et al. 31 kinds; Ni, Cr, W, Mo, Co, Mn, V

Environmental pollution in the century

Environmental quality standards for contamination and monitoring substances:

Cd, Pb, Cr, As, Hg, Se, B, Cu, Zn, Ni, Mo, Sb, Mn

Problem of Rerametals

Depleting resource a few resource, expensive in market

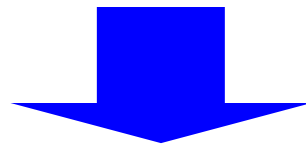
In : 6.2-folds、 Mo : 4,1-folds、 Sb : 2.4-folds increased
for 5 years (2000~2005)

Environmental pollution new environmental quality
standards for contamination

ex)

molybdenum (Mo)	< 0.07 mg/l
antimony (Sb)	< 0.02 mg/l
uranium (U)	< 0.002 mg/l

From 2006 in Japan



Remove and Recover from Environment

Remediation - Recover works

Physical and chemical treatment (coagulation-sedimentation, ion exchanger)

merit : high efficiency from high conc. contaminant, short time

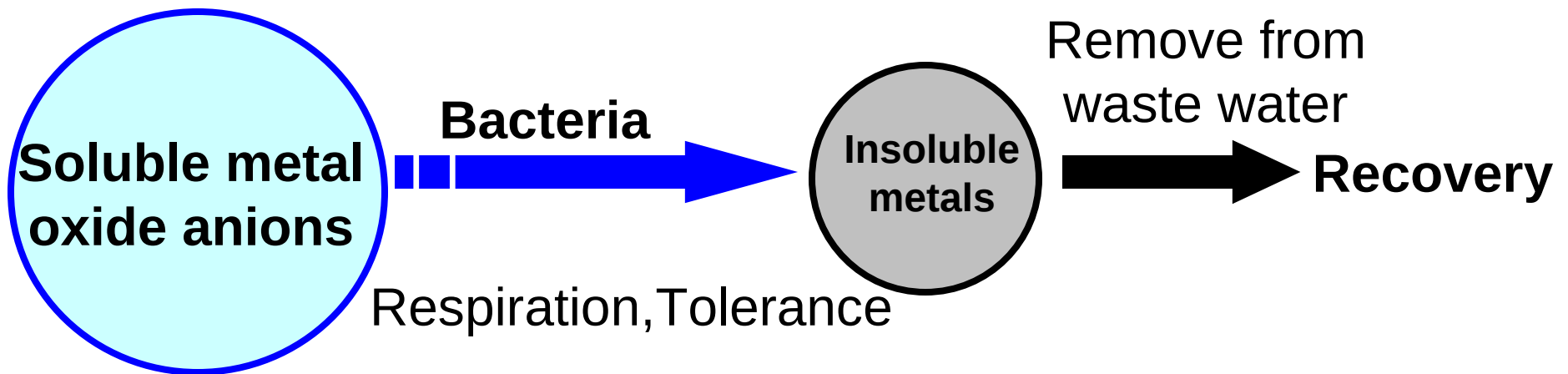
demerit : expensive

Biological treatment (bioremediation)

merit : high efficiency from low conc. contaminant, low cost

demerit : long time

Focus on Bacteria Metabolizing Metals

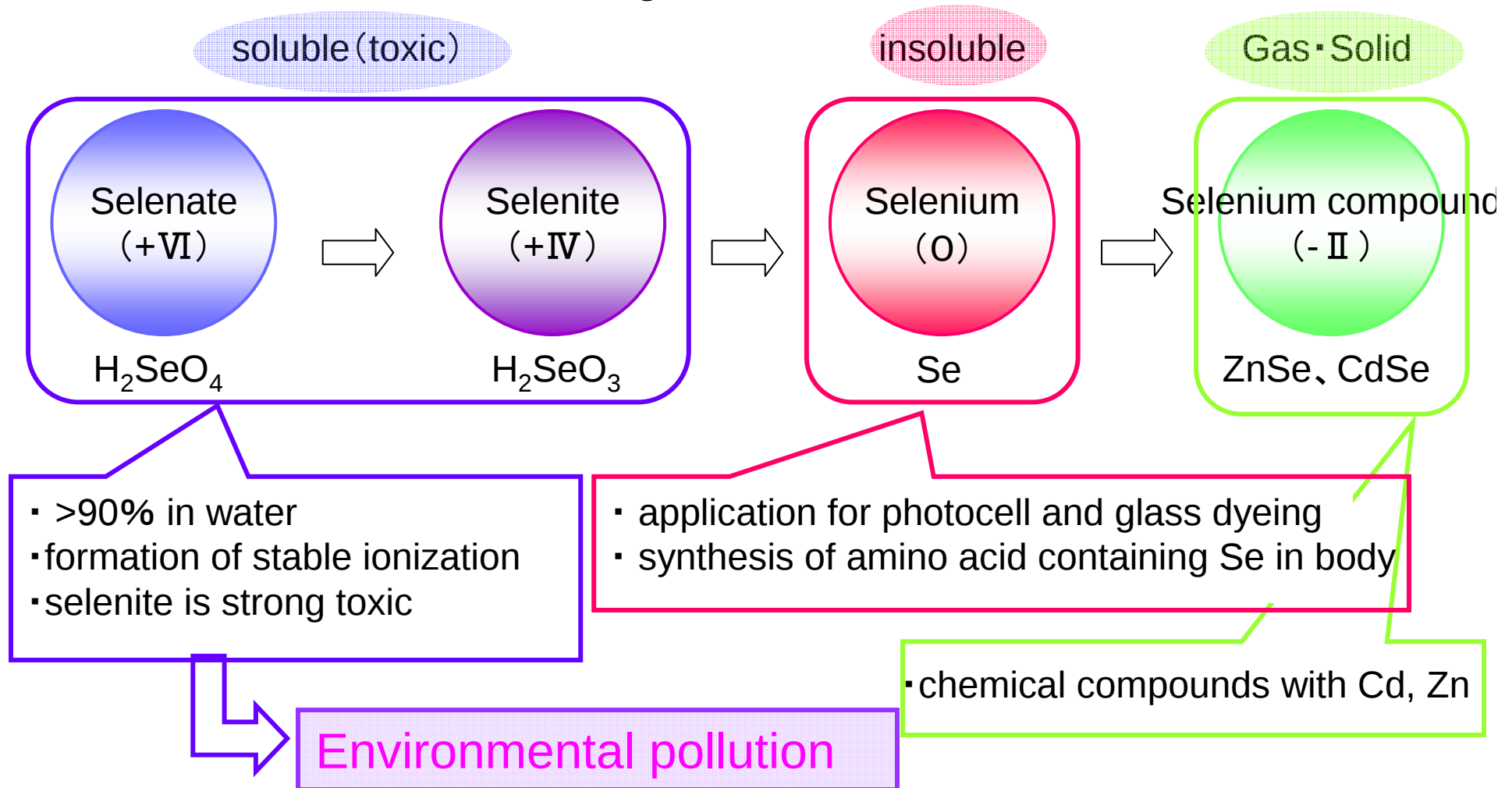


Purpose

- Screening and isolation bacteria available metabolizing rerametals (Se,Te et al.) and recover them
- Analysis the genes and enzymes related to metabolite rerametals
- Construction of platform to enlarge rerametals recover process

Selenium(Se)

- Application for electronic part ➡ **possibility of depleting resource**
- Essential element for organism



Selenium (Se)

Bacillus sp. SF-1

- Isolation from selenium-contaminated sediment (M. Fujita et al., 1997)
- selenate $\Rightarrow$ selenite (anaerobic)
selenite $\Rightarrow$ selenium
- High growth rate under aerobic condition
- Enzyme complex of selenate reduction (S. Yamamura et al., 2004)
- - ◆ membrane-bound enzyme
 - ◆ Mo containing enzyme
 - ◆ induction by selenate under anaerobic

No Host-vector system

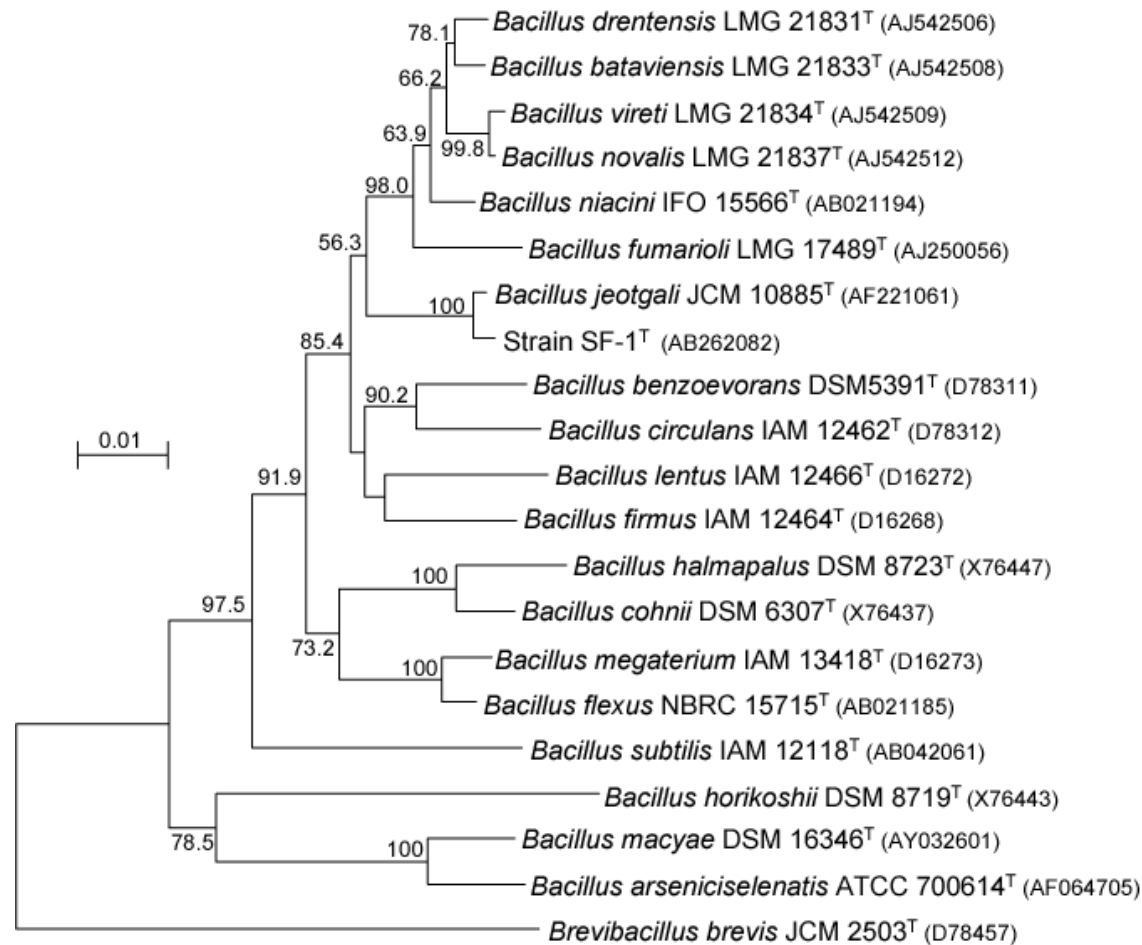


Selenate reduction by SF-1
Cause **red** color

**Purpose $\Rightarrow$ To construct Se remove process by SF-1,
molecular mechanism of selenate reduction is examined.**

Phylogenetic trees of 16S rRNA between *Bacillus* sp. SF-1 and related *Bacillus* species

16S rDNA of SF-1 is 99.6% similar to that of *Bacillus jeotgali* JCM 10885^T



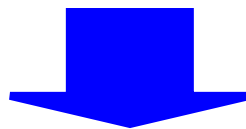
Phenotype between strain SF-1 and JCM 10885^T

Different phenotype in selenate and arsenate reduction

	Strain SF-1	<i>B. jeotgali</i> JCM10885 ^T
Selenate reduction	+	—
Arsenate reduction	+	—
Gram stain	Positive	Variable
7% NaCl tolerance	—	+

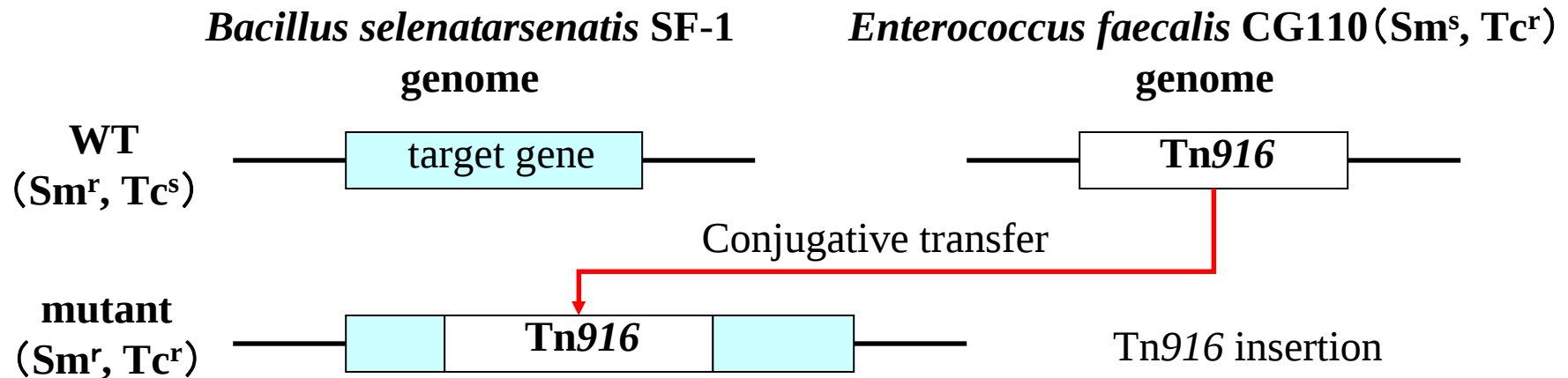
DNA-DNA reassociation value, 14% between SF-1 and JCM 10885^T

Novel species



Bacillus selenatarsenatis SF-1

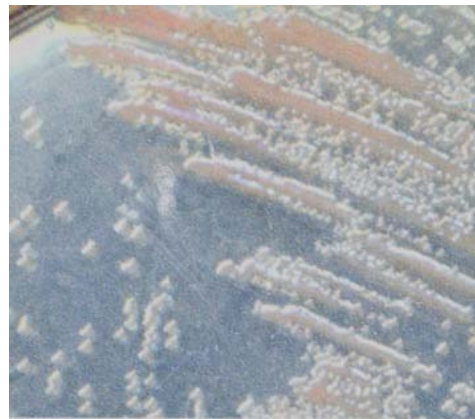
Screening of selenium synthesis-defective mutants



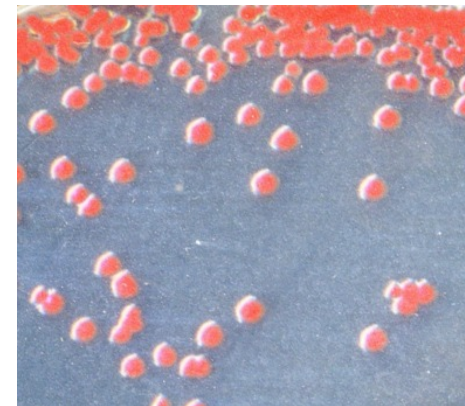
Selenium synthesis-defective mutants
7 colonies (white)



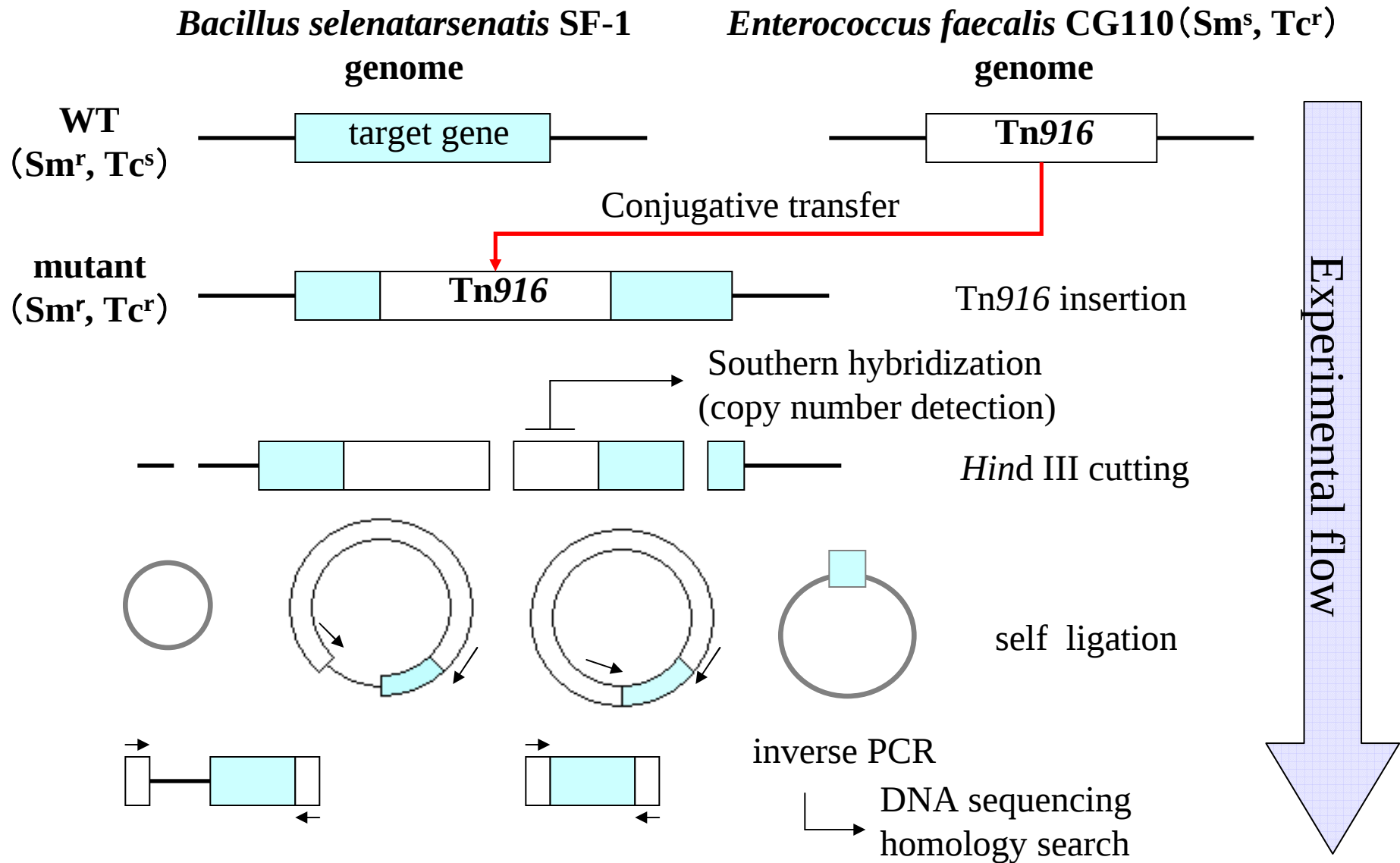
Selenium synthesis-leaky mutants
10 colonies (pink)



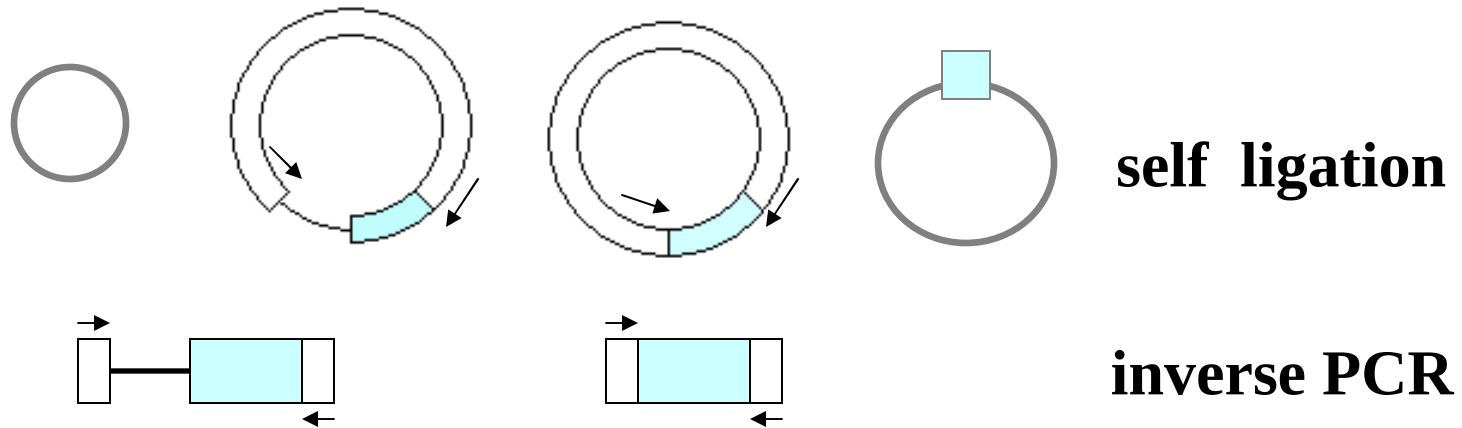
Wild-type (red)



Cloning of target genes using a conjugating transposon Tn916



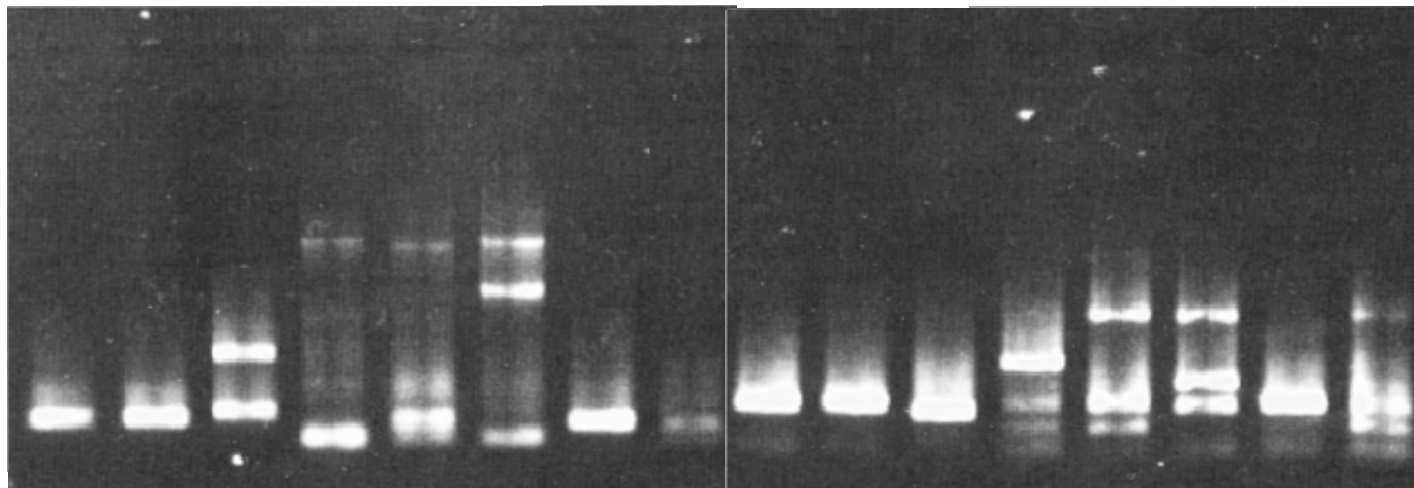
inverse PCR



Upstream Tn916 insertion

Downstream Tn916 insertion

4 11 14 19 23 24 27 WT 4 11 14 19 23 24 27 WT



←4kbp
←2kbp
←0.5kbp

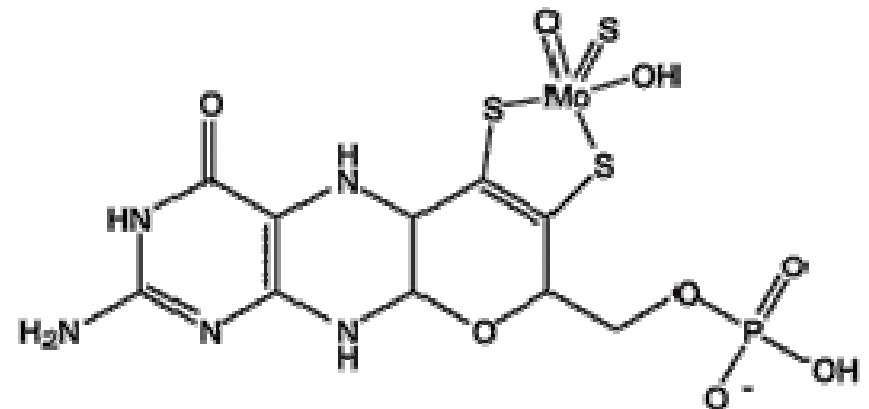
Homology search

- Molybdopterin oxidoreductase
- Diguanylate cyclase / phosphodiesterase (GGDEF domain)

From mutants carrying one copy of Tn916

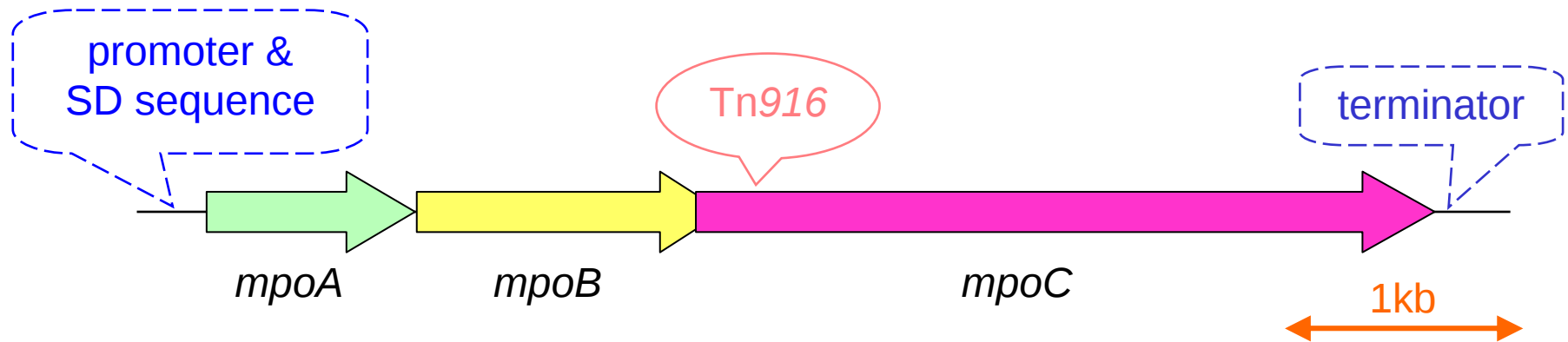
Molybdopterin oxidoreductase

- Generic name for oxidoreductase containing Mo, pterin, Fe-S cluster
- Oxidoreductase related in anaerobic respiration (ex nitrate reductase)



molybdopterin

Analysis of the Genes coding Molybdopterin Oxidoreductase



molybdopterin oxidoreductase,
iron-sulfur binding subunit → ***mpoA*** (0.9 kb)

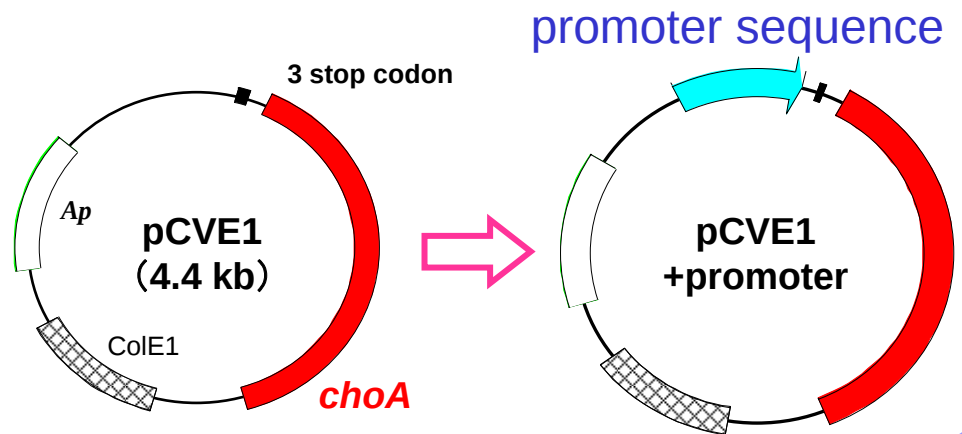
molybdopterin oxidoreductase
membrane subunit → ***mpoB*** (1.3 kb)

molybdopterin
dinucleotide-binding region → ***mpoC*** (3.2 kb)

Identification of Promoter region

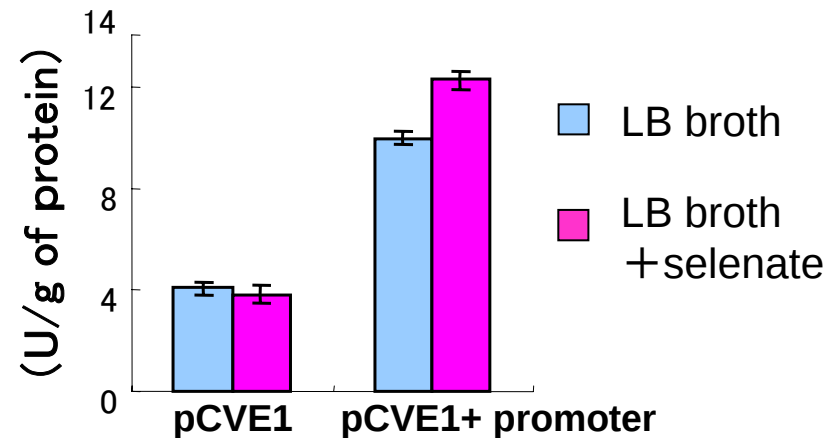
pCVE1

Promoter proving vector with cholesterol oxidase gene (*choA*) as a reporter gene



ChoA assay

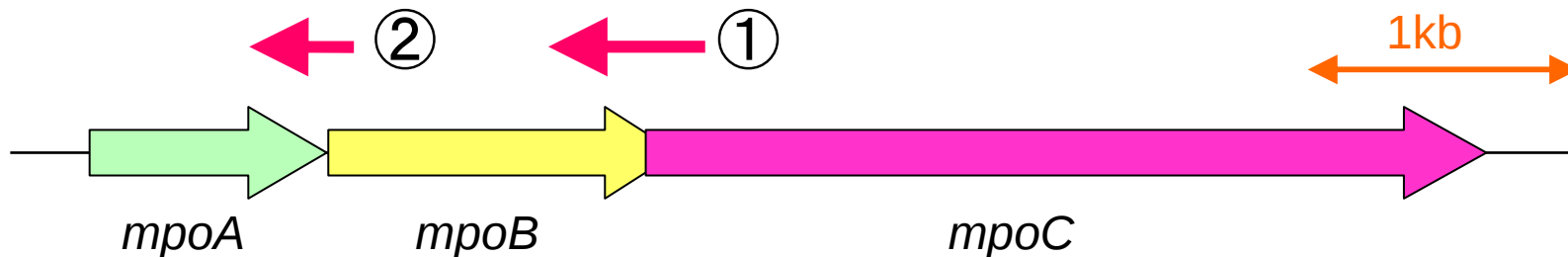
- ① Transform into *E. coli*
- ② Culture • Cell extracts
- ③ ChoA assay with supernatant



Upstream region *mpoA* is promoter

Identification of *mpoABC* operon

Analysis of Transcripts by RT-PCR

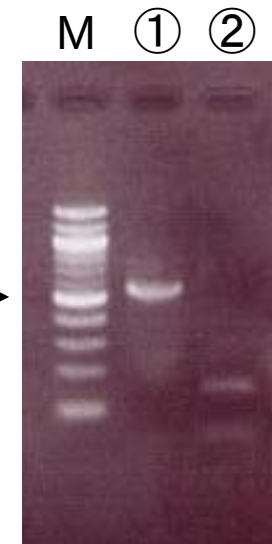


① RNA extracts from SF-1 cells

② cDNA synthesis

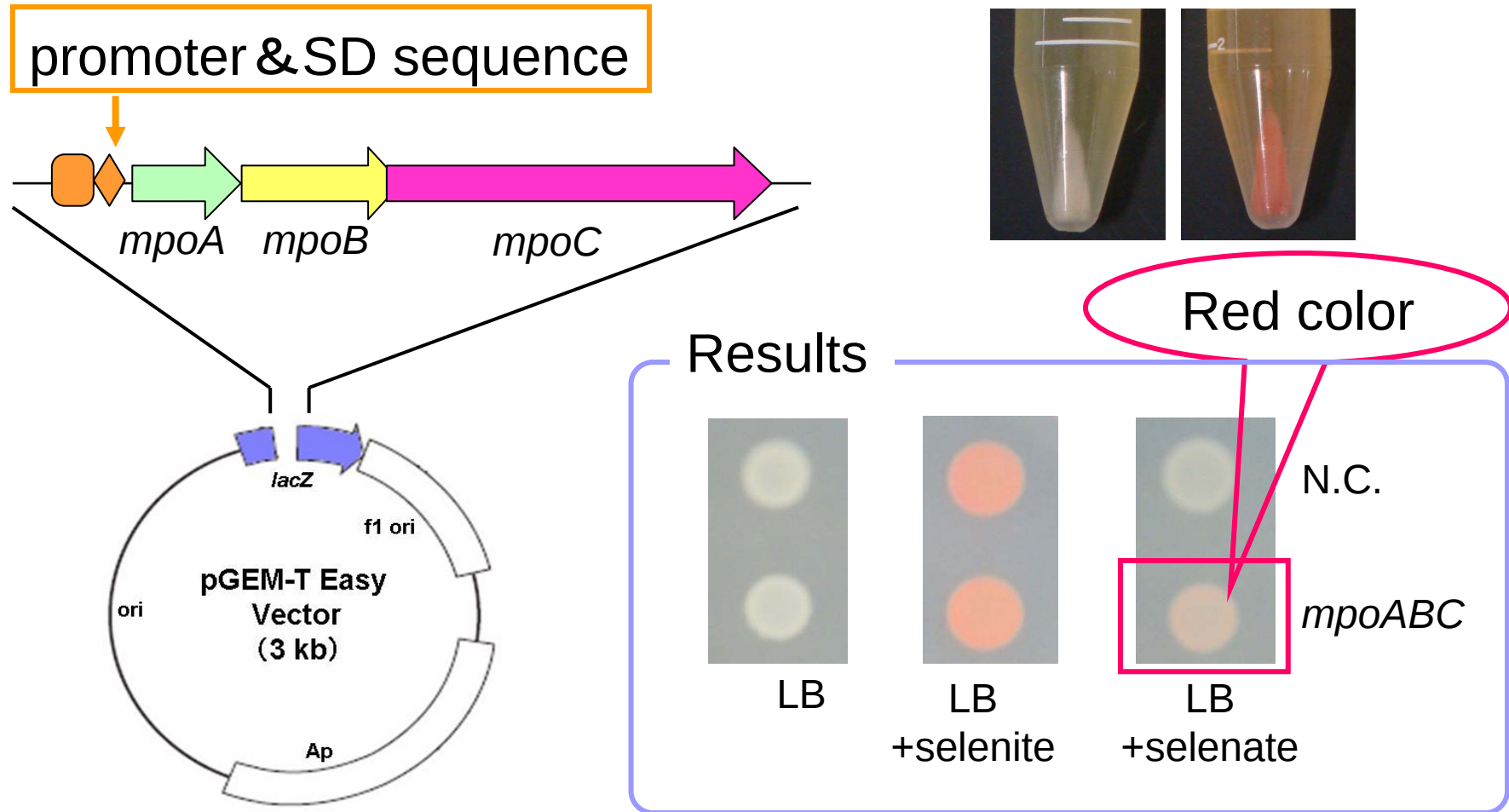
③ DNA Amplification by PCR

0.5kb →



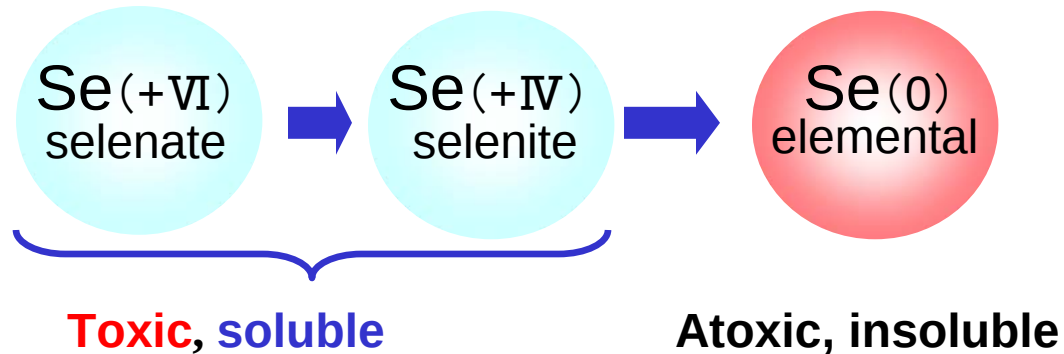
mpoABC is operon

Selenate reduction in *E. coli* carrying *mpoABC*



mpoABC are selenate reducing genes

Back ground Reduction process of selenate by microorganisms



■ Anaerobic condition . . . terminal electron acceptor

■ Aerobic condition . . . detoxification

Merit of aerobe

- easy works (culture • treatment)
- high reaction rate and remove for short time

**Construction of
Aerobic waste
water treatment?**

Few reports of aerobic selenate reducing bacteria

Purpose

Construction of new selenate remediation system

Isolation of novel aerobic selenate high-reducing bacteria and characterization of reduction

Examination Items

- isolation and identification of selenate reducing bacteria
- physiology test
- characterization of reduction (growing cell)

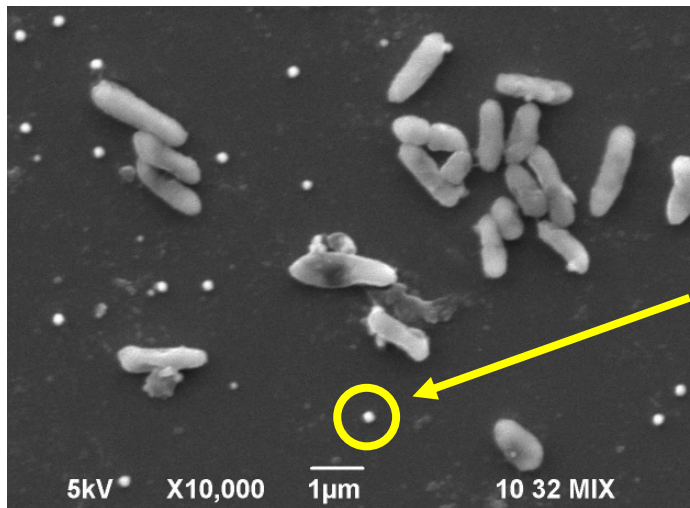
Experiment

Isolation

- Isolation of selenate reducing bacteria, NT-I from an effluent drain of a selenium recycle factory
- SEM and elemental analysis of selenate reducing NT-I and SF-1

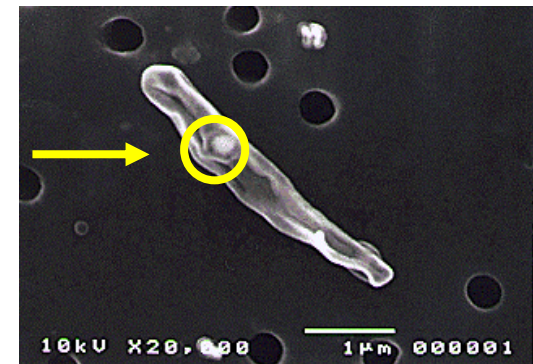


Selenate reducing bacteria



strain **NT-I** (Se is outside of cells)

elemental
Se



strain **SF-1** (Se is inside of cells)

Separation of Se element and NT-I cells by filtration

➡ **Effective recovery of Se element**

Physiology test

Identification

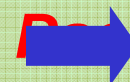
■ Physiology test

cell shape	rod
Gram stain	—
motile	+
O-F test	O
catalase reaction	+
oxidase reaction	+

■ 16S rDNA sequence analysis

100% similarity to that of *Pseudomonas stutzeri* DSM 5190^T

Physiology and Molecular biology test

 ***Pseudomonas stutzeri* NT-I**

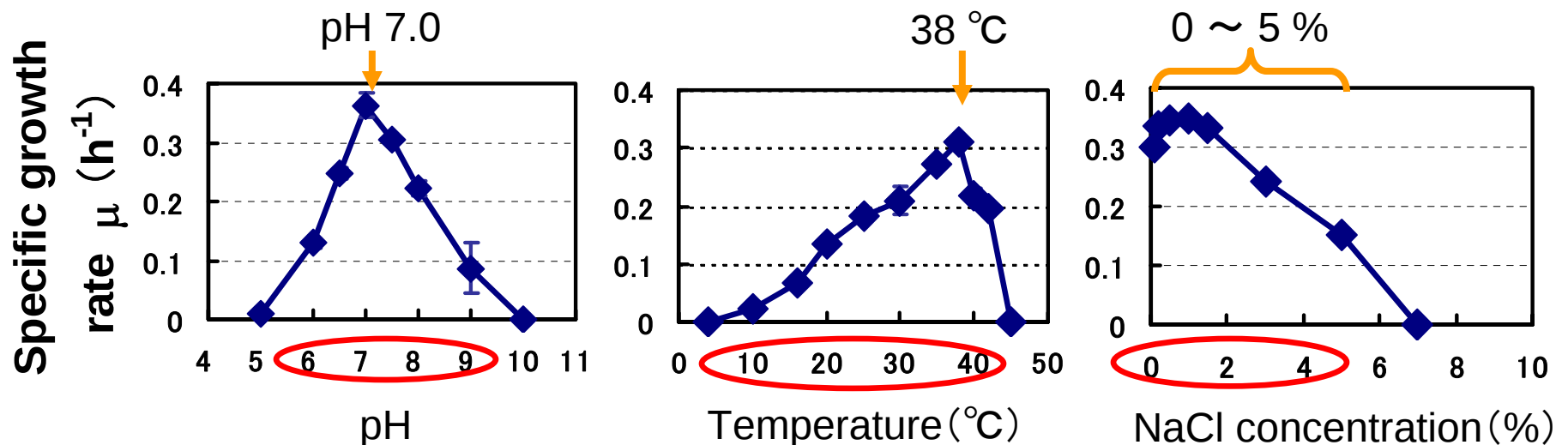
Cell physiology

Physiological characterization of NT-I

■ Carbon Assimilation test (Biolog GN Microplates) → a variety of C source (80 kinds)
(sugar, amino acids, organic acids)

■ Growth rate

Medium . . . Glucose + mineral



Growth: pH 6.0-9.0, Temp. 10-42 $^{\circ}\text{C}$, NaCl tolerance <5 %
Optimum: pH 7.0, Temp. 38 $^{\circ}\text{C}$

Pseudomonas stutzeri NT-I

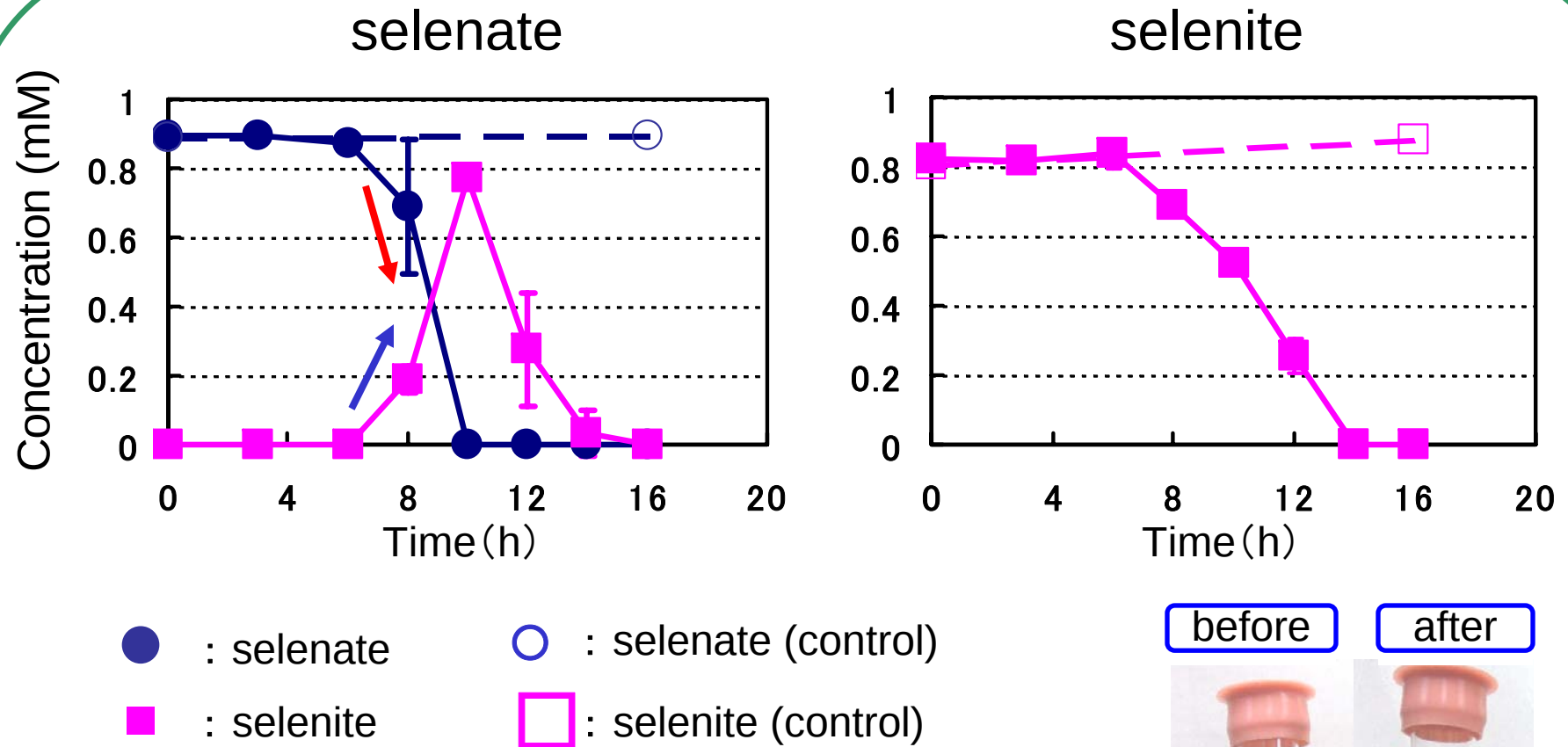
Experiments

Characterization of reduction (growing cells)

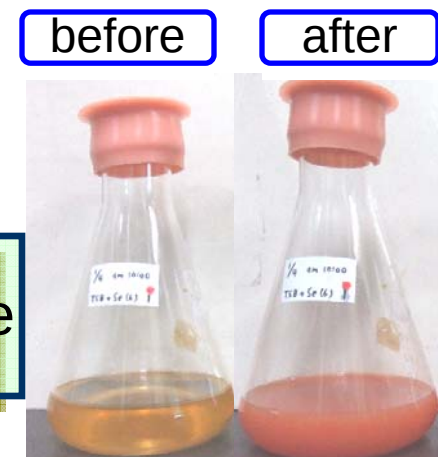
- Medium : TSB + selenate or selenite
- Conditions : pH 7.2, Temp. 28°C, Aerobic
- Comparison : known aerobic selenate reducing bacteria
 - *Pseudomonas stutzeri* ATCC 51152
(Lortie *et al.*, 1992)
 - *Enterobacter cloacae* subsp. *cloacae* SLD1 a-1
(ATCC 700258) (Losi *et al.*, 1997)

Results

Characterization of reduction in NT-I

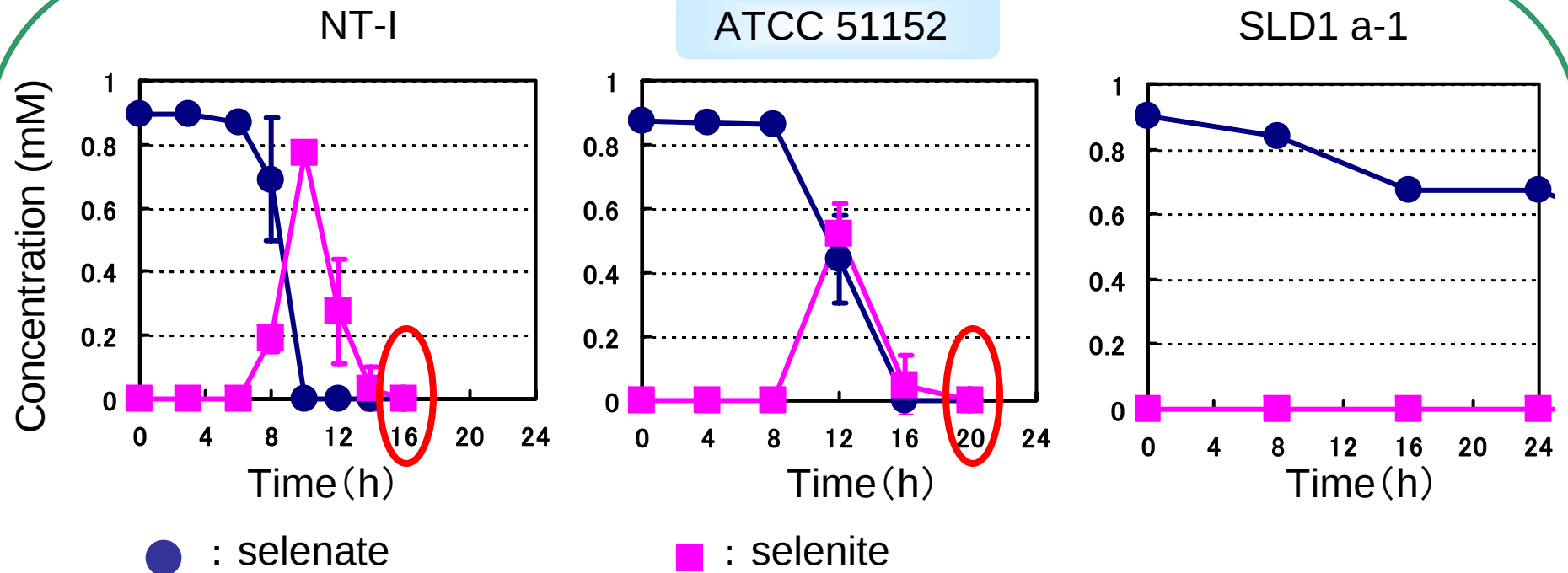


Reduce selenate to Se and Remove from culture



Results

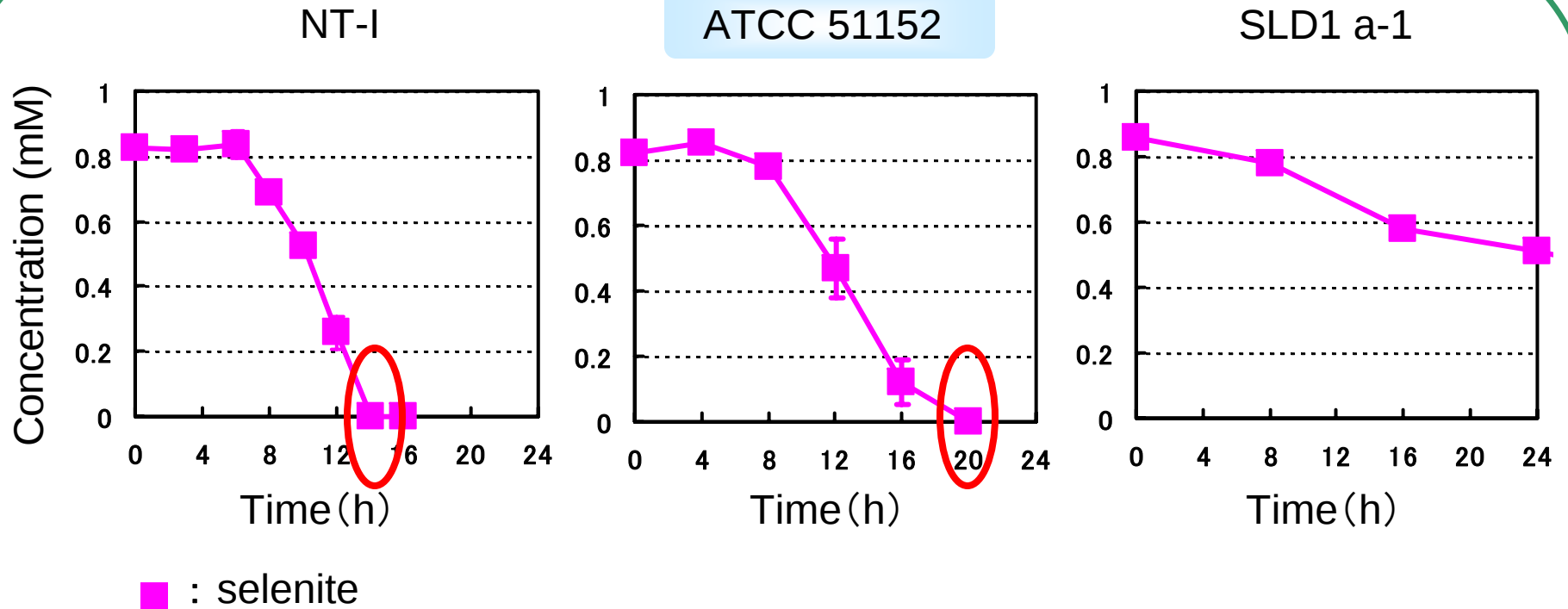
Selenate reduction between three strains



- NT-I ,ATCC 51152 ... 100 % reduction(for 10-16h)
- SLD1 a-1 ... 25 % reduction(for 24h) ,45 % reduction(for 48h)

Results

Selenite reduction between three strains



- NT-I , ATCC 51152 ... 100 % reduction (for 14-20 h)
- SLD1 a-1 ... 40 % reduction (for 24h) , 100 % reduction (for 48h)

Summary

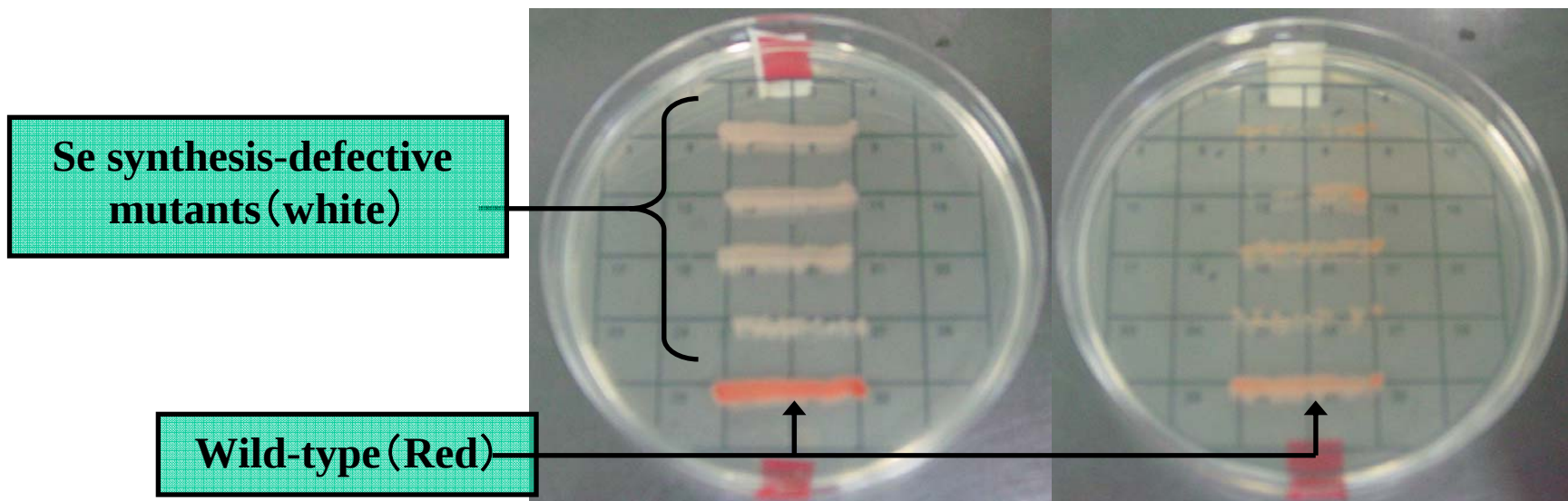
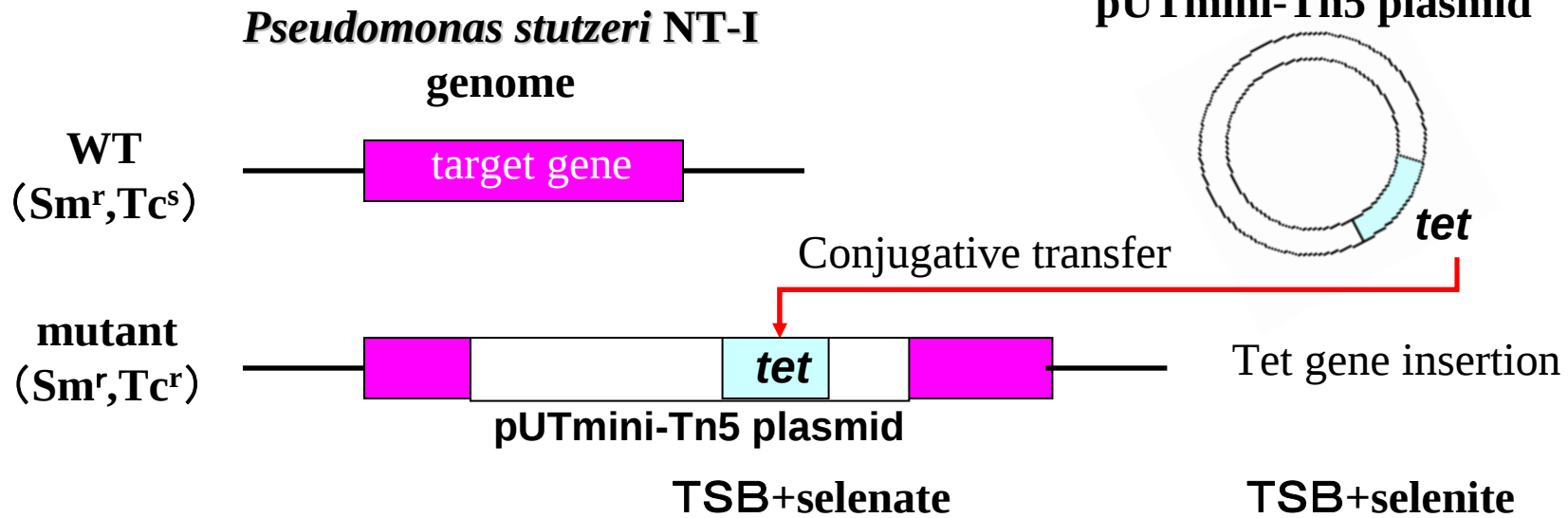
Comparison of reduction between three strains

	ATCC 51152	SLD1 a-1	NT-I
1 mM selenate reduction rate (mol/h)	1.1×10^{-4}	1.2×10^{-5}	2.2×10^{-4}
1 mM selenite reduction rate (mol/h)	8.2×10^{-5}	1.8×10^{-5}	1.1×10^{-4}
maximum selenate reducing concentration	50 mM (Lortie <i>et al.</i> , 1992)	1 mM (Losi <i>et al.</i> , 1997)	≥ 400 mM (visual observation)

NT-I has the most selenate reducing activity

Screening of selenate reduction-defective mutants

E. coli DH5 α (Sm^s, Tc^r)
pUTmini-Tn5 plasmid



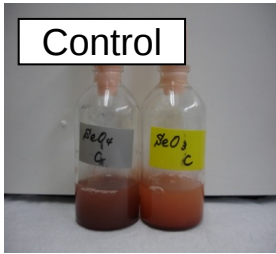
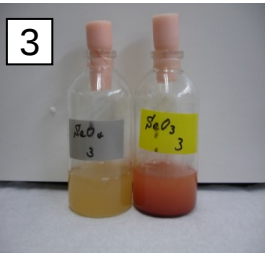
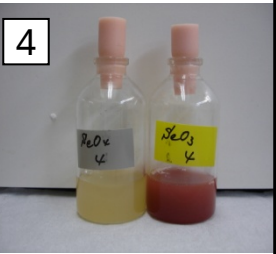
Selenate and Selenite reduction of transposon mutagenesis of NT- I

TSB+H₂SeO₄



TSB+H₂SeO₃

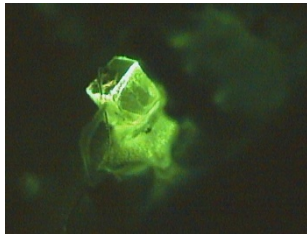


		Control 		1 		2 		3 		4 	
		H ₂ SeO ₄	H ₂ SeO ₃	H ₂ SeO ₄	H ₂ SeO ₃	H ₂ SeO ₄	H ₂ SeO ₃	H ₂ SeO ₄	H ₂ SeO ₃	H ₂ SeO ₄	H ₂ SeO ₃
Conversion to Se (%)	12h	99	33	3.2	34	2.3	40	0.0	37	2.9	39
	24h	99	74	1.1	91	2.4	82	1.4	75	5.9	95

Definite conversion as 0 % from selenate or selenite at 5 mM in initial time to Se.

Tellurium (Te)

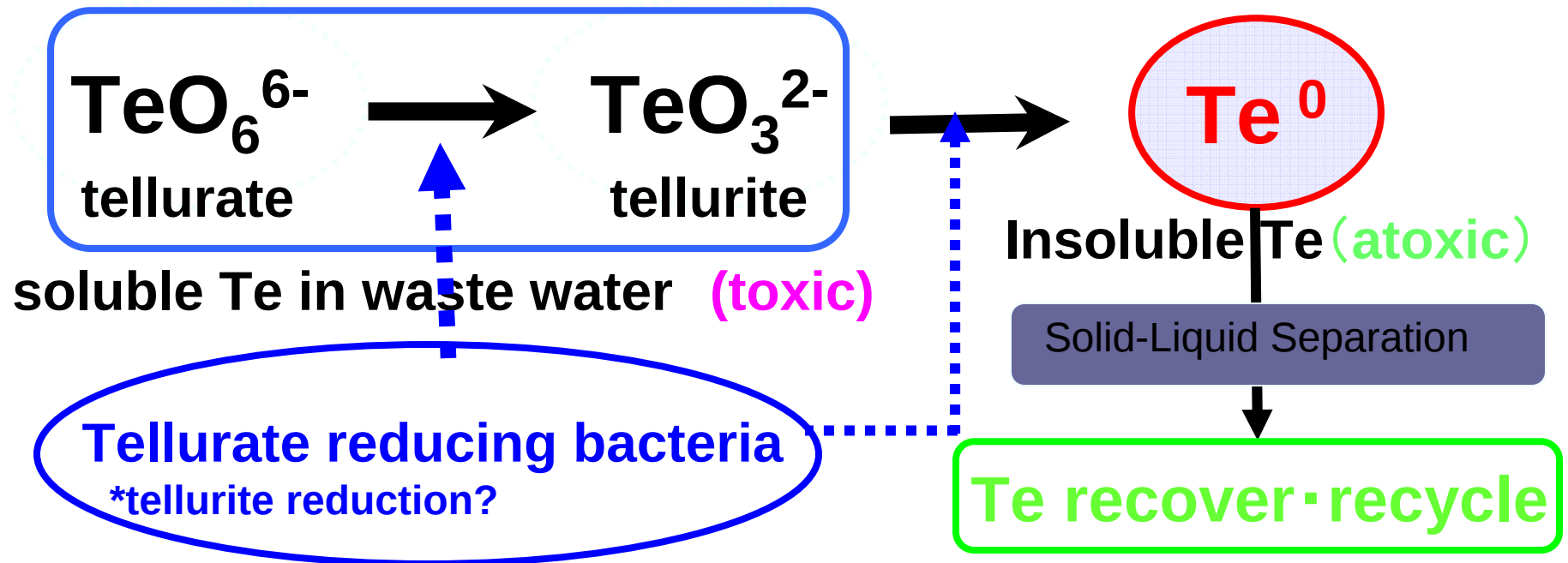
- 16 family element as Se and application of semiconductor material and glass-dyeing
- Soluble oxide and Toxic
- Environmental quality standards for monitoring substances (in Japan)



ZnTe green Light Emitting Diode (LED)



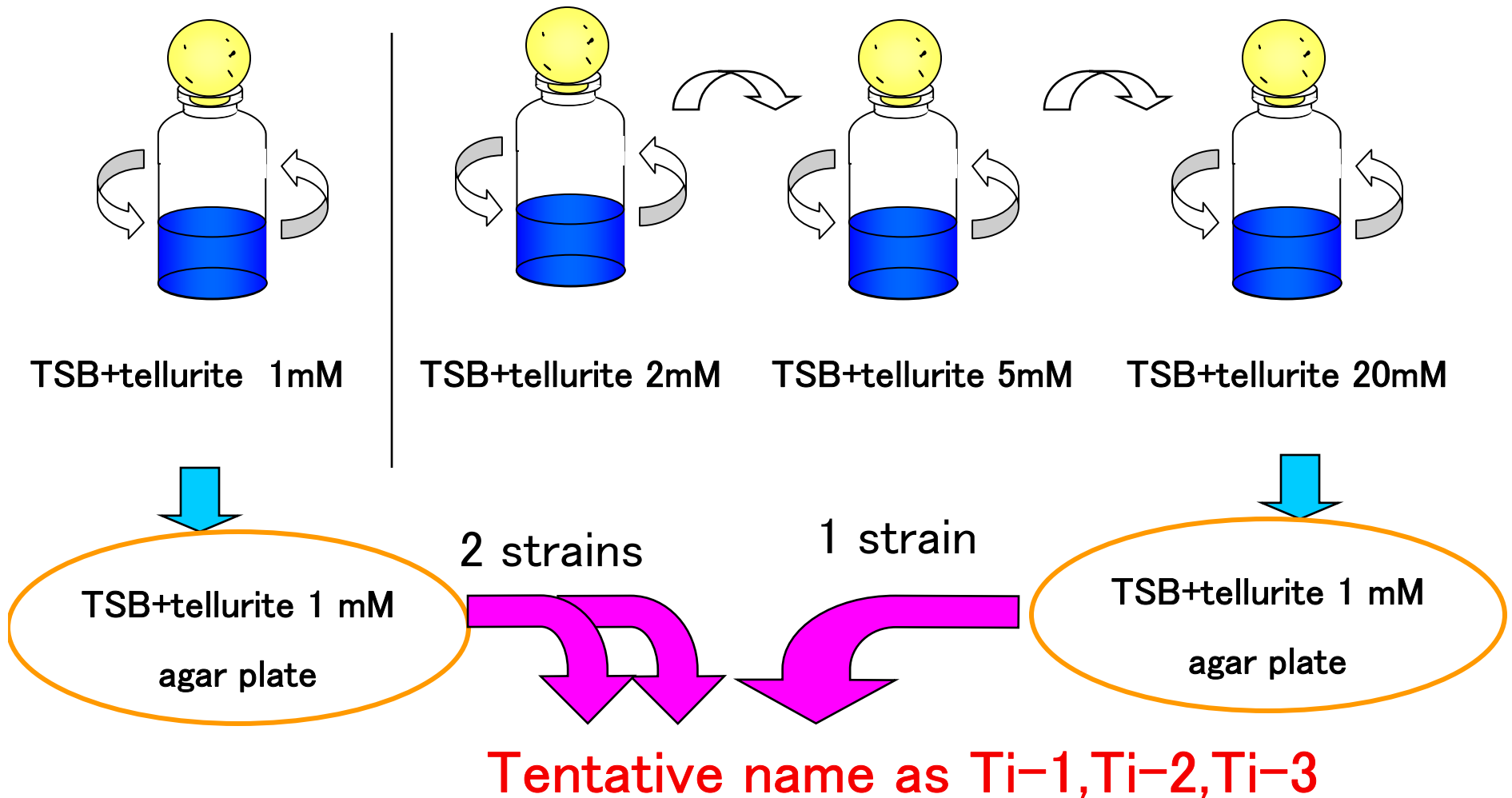
Te elemental mineral: native Te



Isolation of Tellurate Reducing bacteria

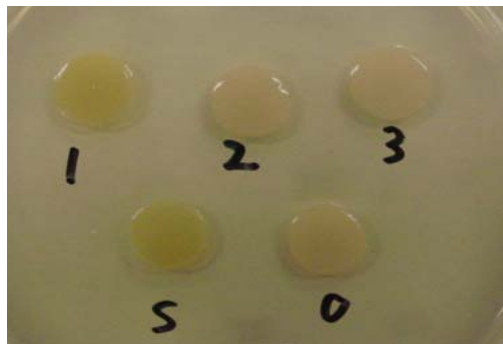
Three strains were isolated from enrichments with tellurite.

Enrichment culture

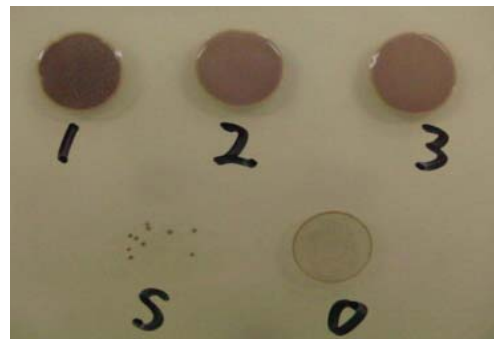


Physiology and Molecular Biology Test in Ti-1, 2, 3

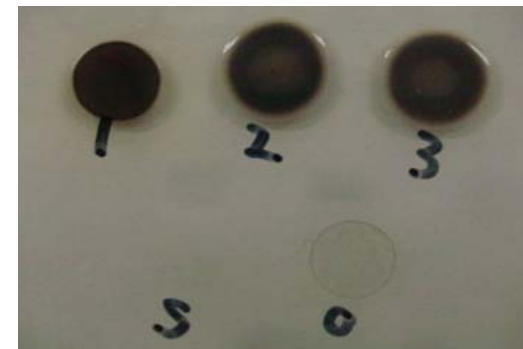
	Ti-1	Ti-2	Ti-3
Cell shape	rod	rod	rod
Gram stain	Negative	Negative	Negative
16S rDNA similarity	<i>Stenotorophomonas maltophilia</i> (100%)	<i>Ochrobactrum anthropi</i> (100%)	<i>Ochrobactrum anthropi</i> (100%)



TSB



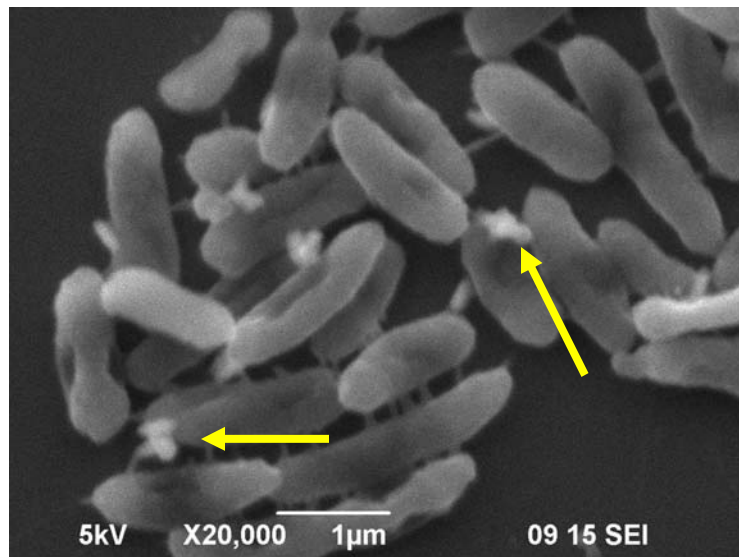
TSB+tellurate1mM



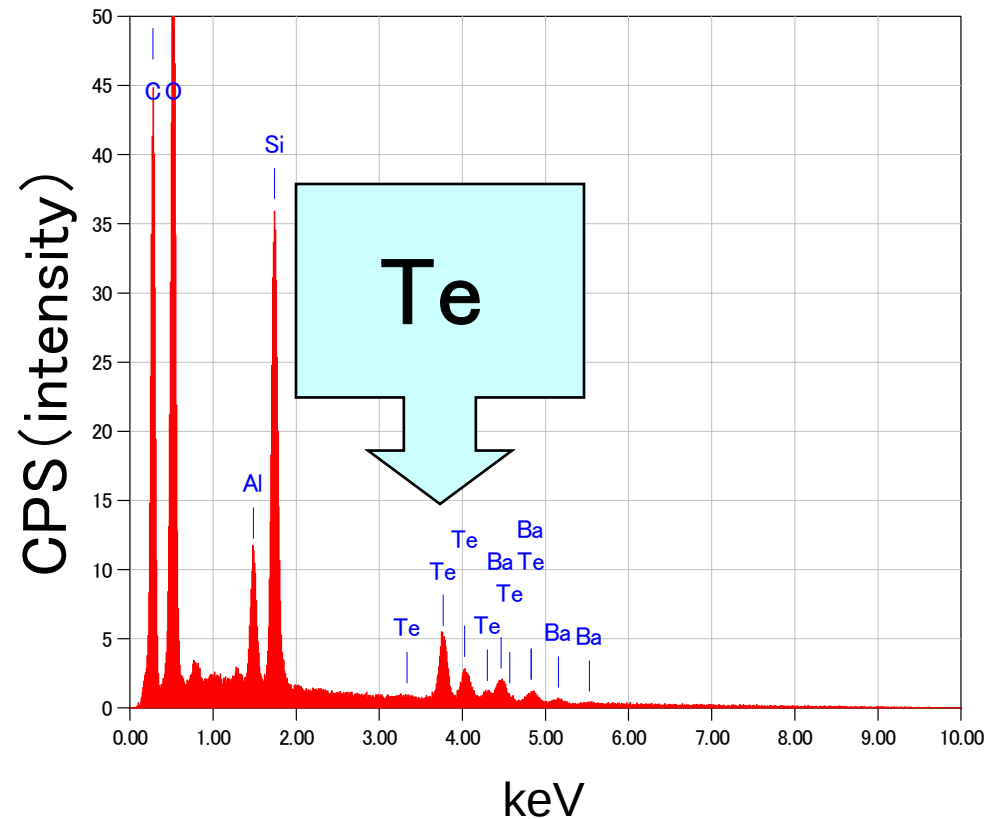
TSB+tellurite10mM

1 : Ti-1, 2 : Ti-2, 3 : Ti-3, S: *Stenotorophomonas maltophilia* NBRC14161^T, O: *Ochrobactrum anthropi* NBRC15819^T

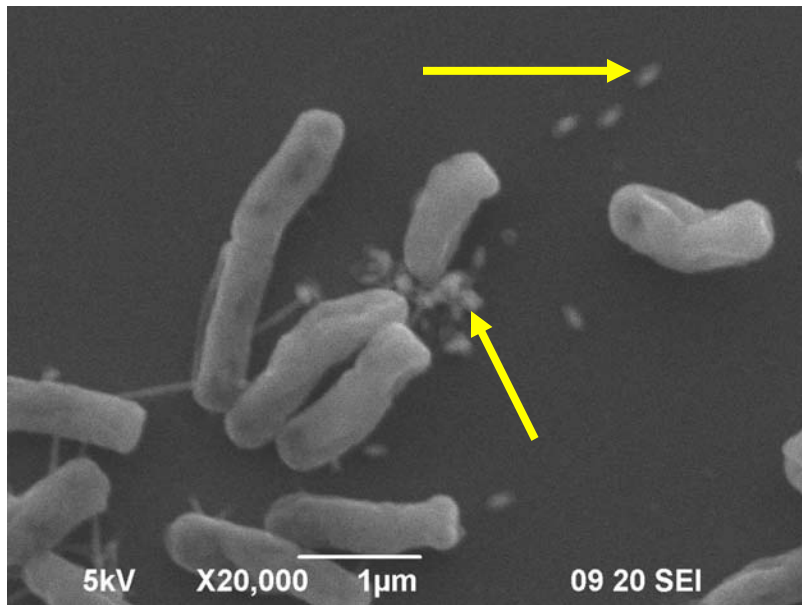
SEM (Scanning Electron Microscope) and elemental analysis in Ti-1



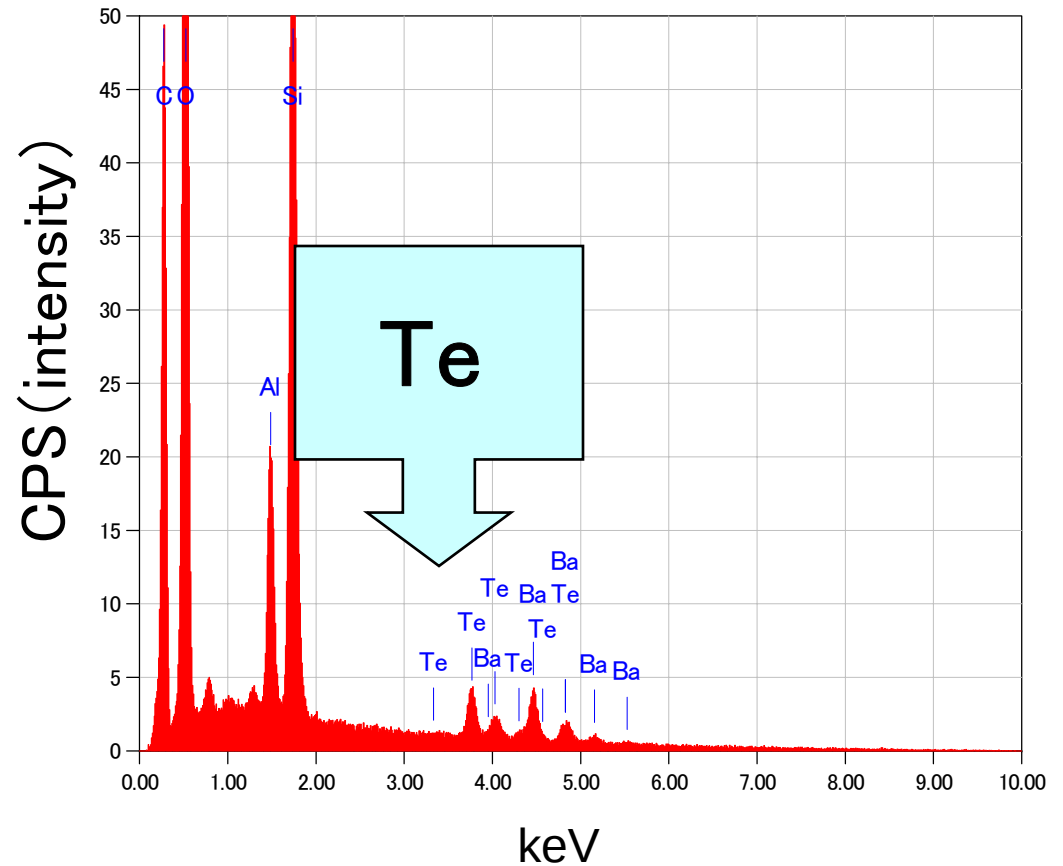
Arrows indicate elemental Te.



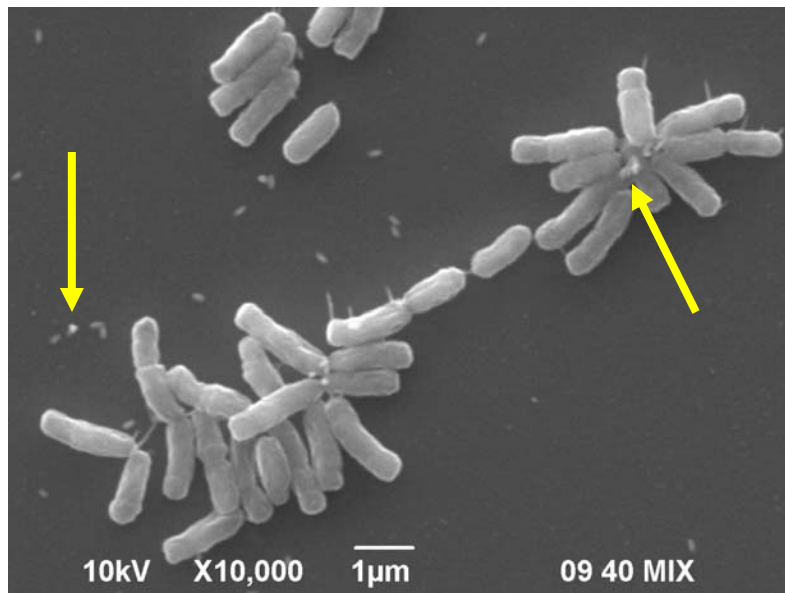
SEM and elemental analysis in Ti-2



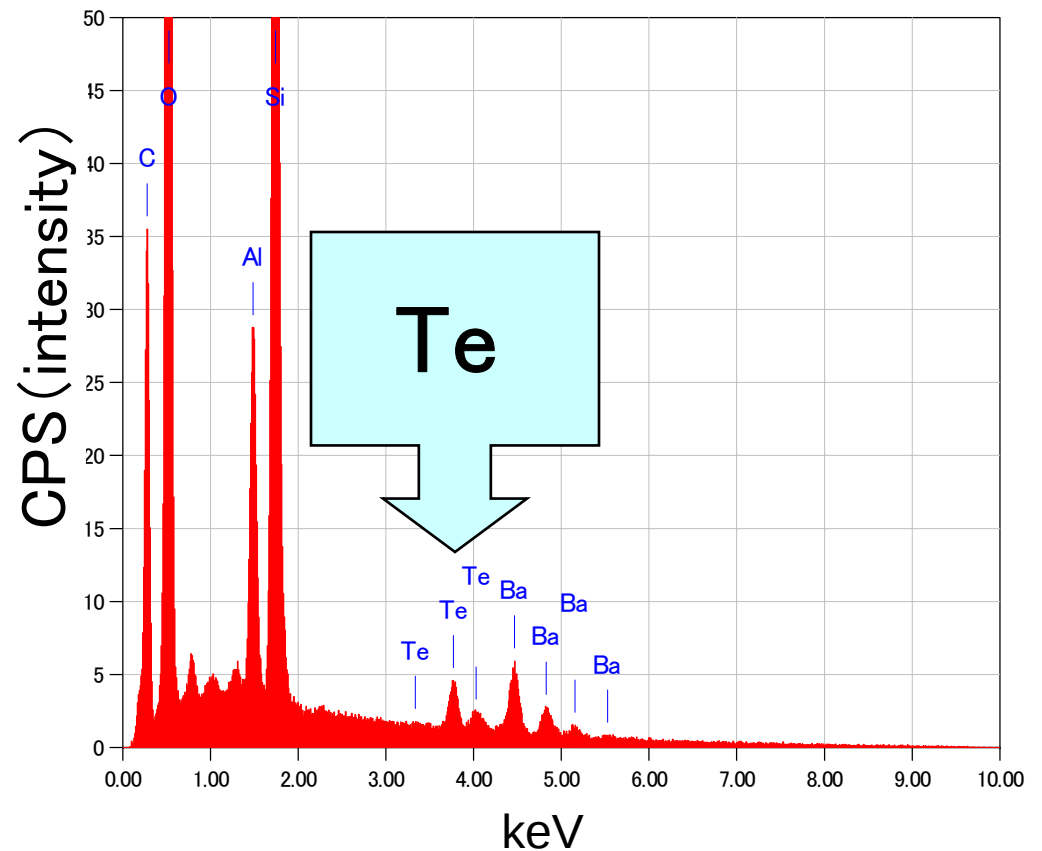
Arrows indicate elemental Te.



SEM and elemental analysis in Ti-3

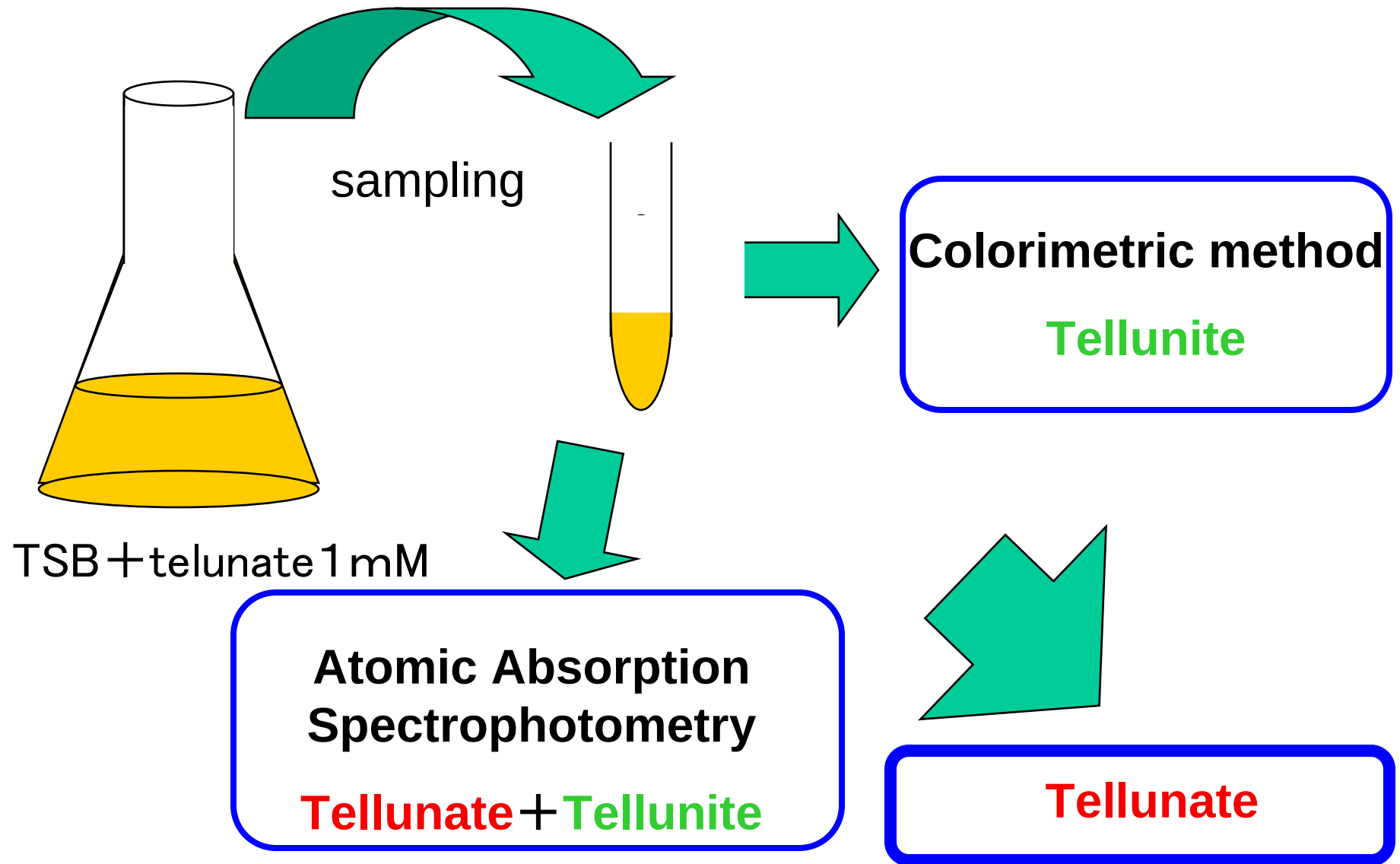


Arrows indicate elemental Te.

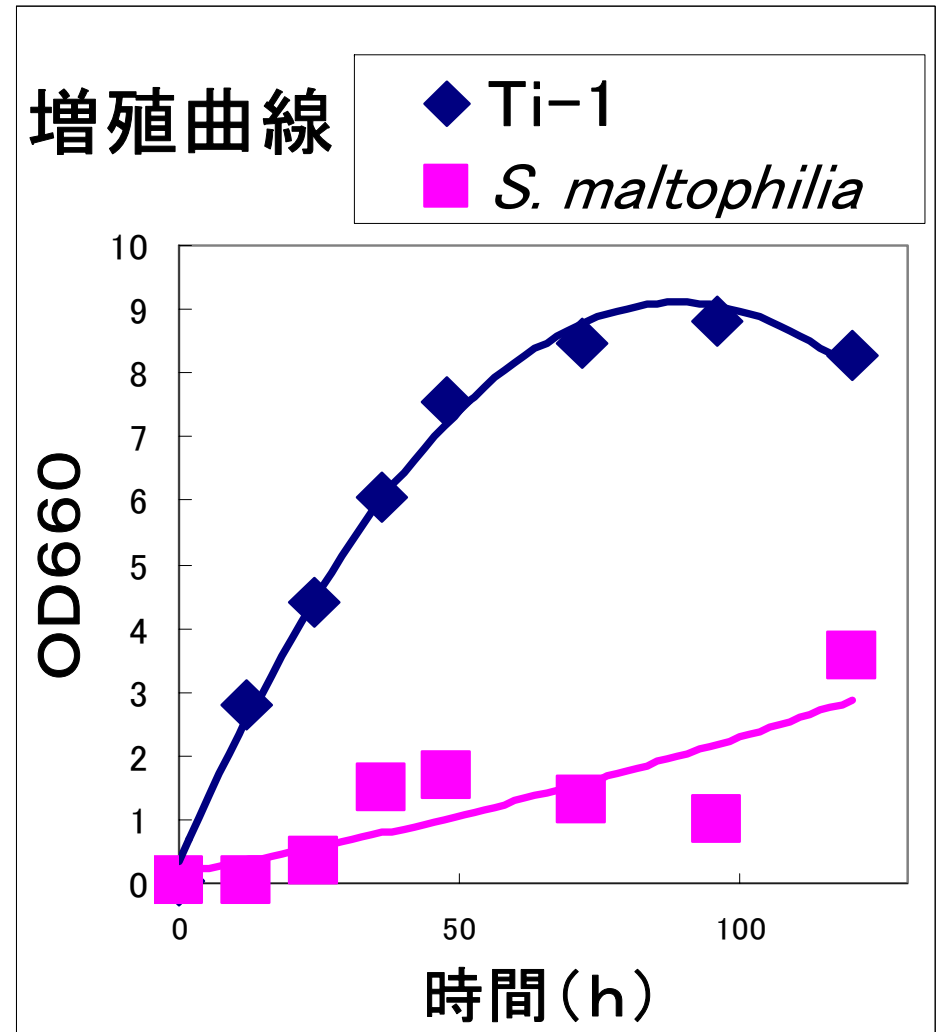
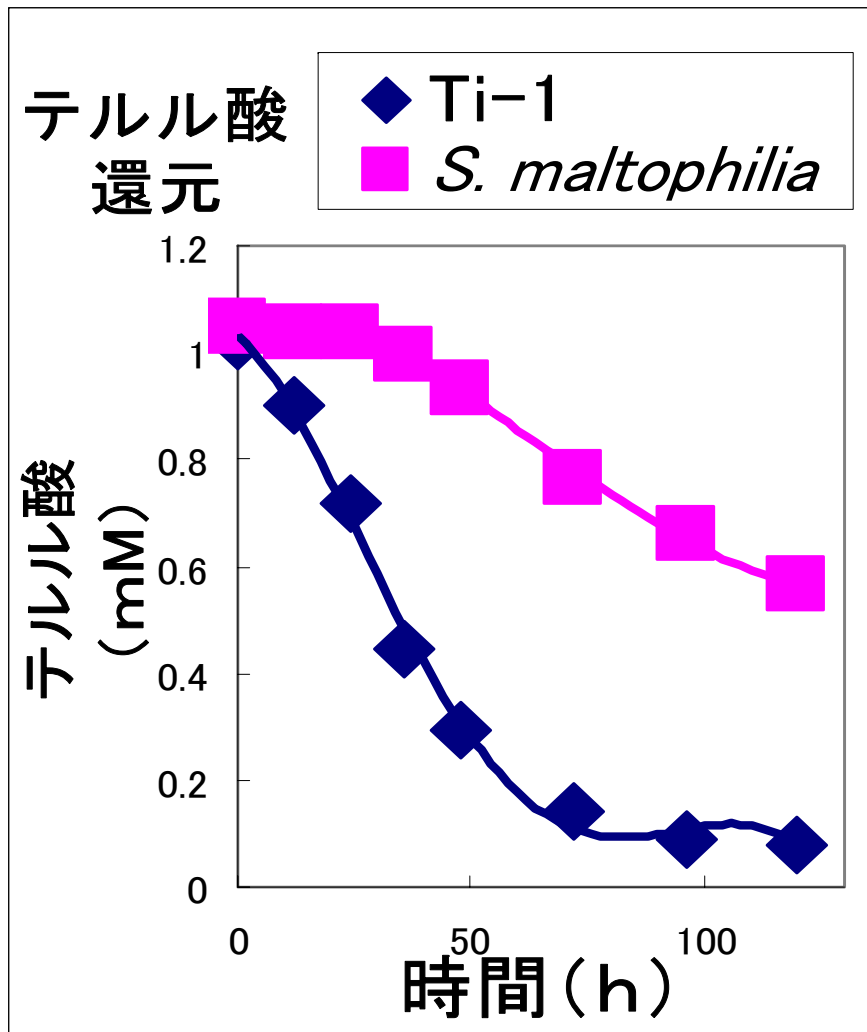


Te element synthesized from Ti-1 are bigger than that from Ti-2, 3.

Measurement of Tellurate reduction

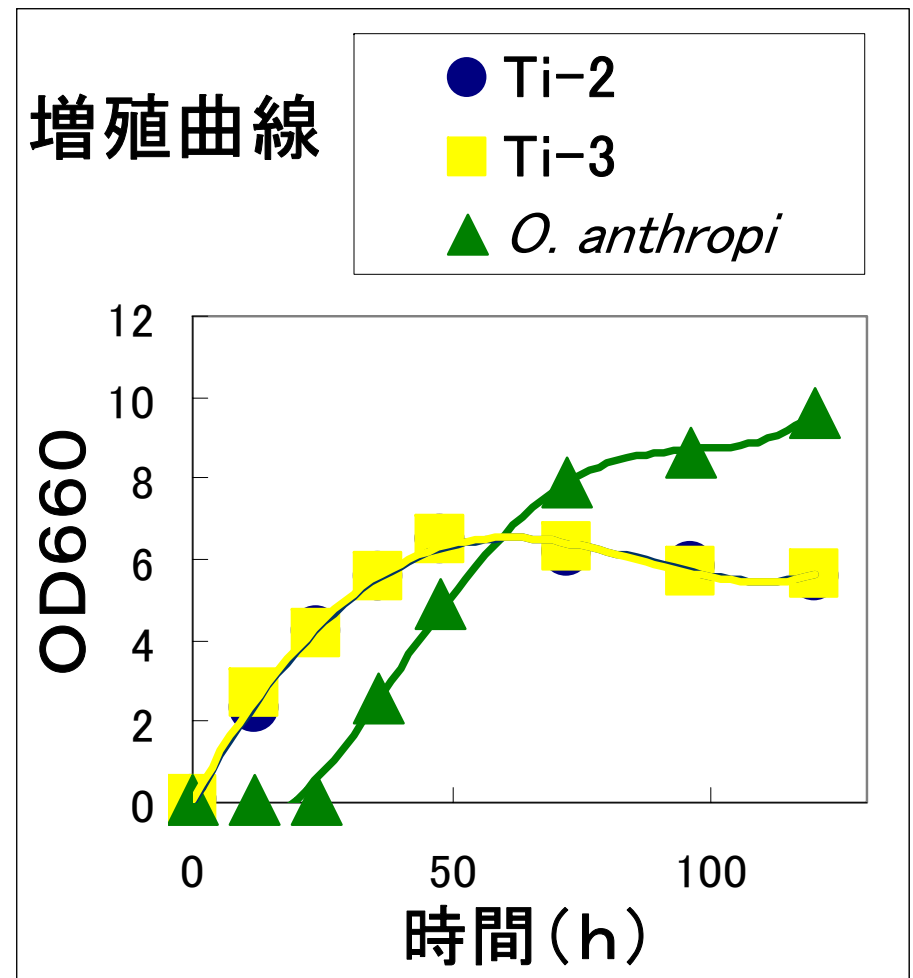
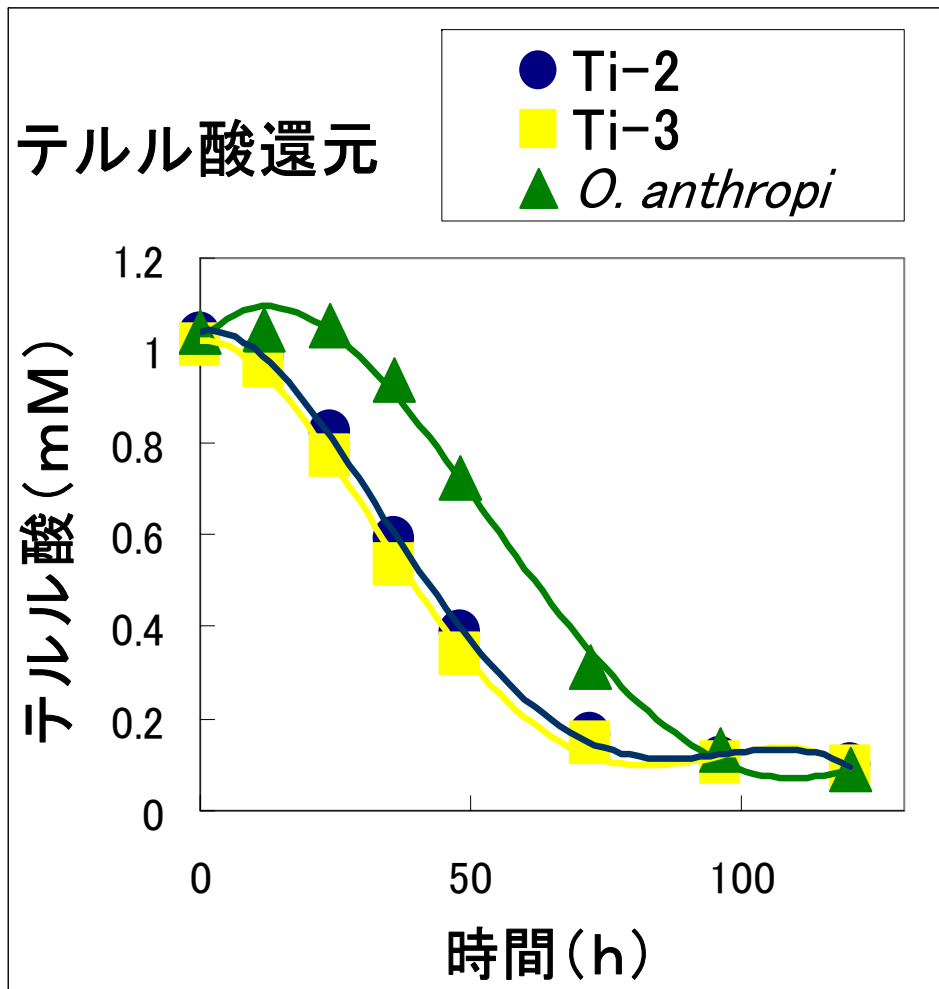


Tellurate reduction between Ti-1 and *S. maltophilia* NBRC14161^T



Ti-1 show high tellurate reducing activity.

Tellurate reduction between Ti-2, 3 and *O. anthropi* NBRC15819^T



Ti-2, 3 show high tellurate reducing activity.

Summary

Selenium (Se)

Identification of SF-1

Cloning of selenate reducing genes

Isolation ▪ Identification as *P. stutzeri* NT-1

High Selenate reducing activity

Tellurium (Te)

Isolation of Ti-1, 2, 3

High Tellurate reducing activity

→ Se and Te Remove and Recycle